

## Review Article

# *Mycobacterium avium* subspecies *paratuberculosis* as a Foodborne Pathogen of High Public Health Significance

Kundan Kumar Chaubey, Shoor Vir Singh

Department of Biotechnology, GLA University, Mathura, Uttar Pradesh, India

## ABSTRACT

*Mycobacterium avium* subspecies *paratuberculosis* (MAP) has potential zoonotic significance and is high risk infection for public health worldwide. It has broad host range and emerged as most successful pathogen of animals and human beings in last 3-4 decades. Animals infected with Johne's disease (JD) shed viable MAP in their milk, fecal, blood, and tissues. Transmission to human beings may occur directly via consumption of animal derived foods, through contact, handling and sharing of space with infected animals and indirectly through contaminated environment and consumption of milk and milk products made from pasteurized milk. Our study on 24,944 animals sampled in last 30 years has shown that MAP is endemic in the livestock population of the country (Cattle-43.5%, Buffaloes-28.6%, goats-21.5% and sheep-35.6%) using 4 diagnostic tests (microscopy, culture, ELISA and PCR). In another study after screening 27,703 human samples (24,322 serum, 3224 blood, 157 stool samples), we found that 34.1, 8.7 and 21.0% of human population was infected with MAP by indigenous serum ELISA, IS900 blood PCR and stool microscopy, respectively. In the absence of control programs bio-load of MAP is increasing with each passing day and if we take total population of animals (500 million plus) and human beings (1.23 billion plus) into account, infected population will go into millions. MAP is not inactivated during pasteurization therefore live bacilli are present in milk and milk products made from pasteurized milk. Presence of live MAP bacilli has also been reported from meat and meat products, soil, domestic water supplies, as well as river water. This article summarizes bio-incidences of MAP in environment, water supplies & aerosols, milk, meat and their products in India vis a vis global status. Besides discussing risk associated with presence of MAP in food chain and impact on human health, paper highlights the knowledge gap between Indian studies vis a vis International scenario.

**Keywords:** Paratuberculosis; *Mycobacterium avium* subsp. *paratuberculosis*; Food; public health; zoonotic; foodborne pathogen

## Introduction

*Mycobacterium avium* subspecies *paratuberculosis* (MAP) is a major animal pathogen which has inflicted huge losses to the livestock industry worldwide (Vinodhkumar et al., 2013; Rawat et al., 2014; Bauman et al., 2016). It is very difficult to diagnose the MAP infection in the early stage of the infection. It has long been known and well established that animals infected with JD (clinical and sub-clinical), shed MAP bacilli in their milk and feces (Streeter et al. 1995;

Raghuvanshi et al., 2013). Contamination in the milking parlor is one of the commonest source of contaminating milk with MAP (Nacy and Buckley, 2008). MAP is regarded as important food borne pathogen and has been reported to infect wide range of animal species, including non-human primates and human beings (Waddell et al., 2008; Kumar et al., 2010; Naser et al., 2009; KuKanich et al., 2013; Hussain et al., 2015). It may colonizes in animals for years without emerging clinical disease. Sub-clinically infected animals shed MAP in their milk (Shankar et al., 2010) and feces and contaminate pastures, environment and food chain for long time (Shankar et al., 2010; Singh et al; 2012c). Live MAP bacilli has been also recovered from pasteurized milk (Ellingson et al., 2005), infant formulation made from pasteurized milk products (Hruska et al., 2011),

Received: 10.10.2018, Revised: 08.12.2018,  
Accepted: 20.12.2018

✉ Kundan Kumar Chaubey

E-mail: [kundan.chaubey@gla.ac.in](mailto:kundan.chaubey@gla.ac.in)

surface water and soil samples (Singh et al., 2012c), cow manure ‘‘lagoons’’ that leach into surface water and municipal / tap water (Collins et al., 2003), thus providing MAP multiple routes of transmission to infect human population. Cow manure in solid and liquid forms is applied as fertilizer in agricultural land (Grewal et al., 2006; Gill et al., 2011), plants (upper greens, roots) and soil (surface and depth of 80 cm from plant roots) of agricultural and grazing areas hold MAP for longer period of time (Kaevska et al., 2014). MAP being thermo tolerant survives commercial pasteurization (72°C for 15-30 seconds) temperature (Grant et al., 1998) as well as high salt concentrations (Gao et al., 2002). Filtration, cold shock, hydrostatic pressure and pulsed electric fields procedures have already been tested on milk (Rowan et al. 2001). This superior survival efficiency and dormancy allows the pathogen to be more insidious in human beings (Whittington et al., 2005). Present paper reviews the public health significance of MAP infecting domestic livestock and related issues regarding presence in natural environment, survival abilities, pathogenicity and association with number of human diseases.

#### **Association of MAP and Crohn’s disease**

JD is presently not in focus for the control and eradication either in the veterinary sciences field or in the medical sciences, therefore both animal and human population continue to get widely exposed to MAP infection from sub-clinical and clinical shedders of bacilli (Jones et al., 2006). MAP also has neuropathological and immune dys-regulatory properties (Scanu et al., 2007). This is why MAP is considered a strong candidate pathogen having association with CD (Momotani et al., 2012). Frequent isolation of MAP from CD patients and similarities of complication of CD matched with JD, which further confirmed the involvement of MAP infection with CD (Scanu et al., 2007).

Identification of MAP in CD patients has been accomplished by several different techniques. MAP has been cultured from the intestines ( Sechi et al., 2005) and blood (Naser et al., 2004) of Crohn’s disease patients. In addition, MAP has been identified in the intestines and blood of Crohn’s patients by IS900 PCR amplification ( Naser et al., 2004; Abubakar et al., 2008). IS900 PCR also detected MAP in two tissues of cervical lymph nodes of a 5 years child; later he developed CD (Hermon-Taylor et al., 1998). Multiple serological diagnostics have confirmed the antibody to

MAP in human beings suffering with CD (Feller et al., 2007). CD share certain clinical as well as histopathological similarities with JD and is fast emerging as major disease of public health significance and a potential human zoonotic infection (Singh et al., 2012a). MAP has been detected in the tissues and blood of CD patients with a greater frequency (Naser et al., 2004), human breast milk of a patient with CD, positive antibodies to MAP antigens in blood samples of CD patients as compared to controls (Naser et al., 2000). In-situ DNA hybridization method has allowed direct visualization of small numbers of MAP in CD patient intestines by light microscopy (Abubakar et al., 2008). Initially, Dalziel had characterized the 13 cases of chronic intestinal enteritis where, he examined nine patients histologically and clinically and found that jejunum, transverse and sigmoid colon, as well as mid-ileum were affected (Dalziel, 1989) and patients had similar clinical findings as cattle suffering with JD. He therefore speculated that MAP could be a prospective etiological agent for the complications observed in human patients (Dalziel, 1989). Chiodini et al. (1984) was revived the association of MAP with CD and reported the isolation of un-characterized mycobacteria from tissues of three CD patients and Yoshimura et al. (1987) anticipated that pathogen present in a cell-wall deficient form was characterized as MAP. In a study, MAP was directly visualized by light microscopy in small numbers in the intestines of CD patients by ZN staining (Jeyanathan et al., 2007), while, other pathogen have also been detected in the intestines of patients with CD (De Hertogh et al., 2008). Some researchers argued that MAP has met both Koch’s postulates (Greenstein et al., 2003) and Relman’s criteria (Chamberlin et al., 2007) for microbial causation of CD. The causal association of MAP with CD remains controversial (Miller et al., 1996). Earlier studies from India on random screening of human population from different geographical regions of North India showed 23.4 to 34.0% seroprevalence of MAP (Singh et al., 2011; 2014b). Study reported moderately higher existence of anti-MAP antibodies in human population, which demands urgent measures and programs for reducing the bio-load of MAP in the animal population and in the environment as well.

#### **MAP in water supplies and aerosols**

Large number of studies reported survival of robust zoonotic pathogens in the environment (Szewzyk et

al., 2000). It has been suggested that there is a high risk of transmission of MAP to human population through drinking-water or by aerosols (Hermon-Taylor et al., 2000). Water bodies like lakes and rivers get contaminated by run-off of MAP from heavily grazed planktonic form of pastures, within protozoa or, more likely, both (Pedley et al., 2004). These organisms reaching domestic outlets in high dilution may aggregate in bio-films present in cold and hot household water storage tanks and delivery systems. Mycobacteria are well-known to present in aerosols and to concentrate in the water droplets (Blanchard & Syzdek, 1972; Wendt et al. 1980).

Heavy rain falls from the Atlantic washes off contaminated pastures into rivers in spate. Taff (one of these rivers in spate), goes through the middle of Cardiff city. Research carried out during the 1970s in Cardiff, recorded significant increased incidence of CD ( $p < 0.001$ ), but not of ulcerative colitis, in 11 of the local electoral city wards (Mayberry & Hitchens 1978). Of these high incidence wards, eight directly touched the river Taff and the three that did not, were closely adjacent to the North and East. This is the direction in which aerosols would be carried off by the usual South-Westerly winds (Hermon-Taylor 1993). Tracheal and bronchial inflammation with abnormal lung function tests were noticeable in the significant proportion of human population and chronic granulomatous tracheo-bronchitis in children with CD (Naser et al., 2009; Singh et al., 2012a). These are only few studies but much research is required on MAP presence and survival in the environment, in surface and ground waters and in the aerosols.

### MAP in the natural environment

It is assumed that the survival of *Mycobacterium bovis* in the environment to be limited to hours or days. By contrast, MAP is physically more robust than other mycobacteria and is known to survive for months and perhaps years. Geographical regions having acidic soils enriched with humic and fulvic acids, boreal forests and areas with a heavy rainfall may favor the accumulation and persistence of MAP in the environment (Singh et al., 2012b; Hruska et al., 2012; Thomson et al., 2013; Rhodes et al., 2014). There are no reported studies to give us detailed understanding of the ecology and prevalence and presence of MAP in the atmosphere and potential cycling of MAP through human populations. It is very much possible that environmental MAP is taken up into protozoa and

adapts in intracellular condition within protozoa present in the environment and in biofilm communities that profoundly influence microbial survival, phenotype and virulence. (Barker & Brown, 1994; Hermon-Taylor et al., 2000). MAP grown in vacuoles in *Acanthamoeba castellanii* has developed the capacity of MAP to infect other amoebae, macrophages and human colonic epithelial cells including improved virulence for infection in a beige mouse model (Cirillo et al. 1997). MAP can remain alive for long periods inside the *Acanthamoeba polyphaga* (Steinert et al. 1998). The environmental exposure of MAP and its cycling through unicellular living beings as well as multi-cellular animal and human populations, has potential to have a profound effect on the evolution of these organisms and the evolution of strains with respect to adaptation and enhanced pathogenicity. Previous studies from England, France, Australia and US reported that soil types and soil composition have influence on maintenance, survival and repeated infection and incidence of JD in livestock herds (Dhand et al., 2009). Singh et al. (2012b) screened 71 samples from soil (51) river and water (20) (natural resources) of three districts of South Uttar Pradesh by microscopy and IS900 PCR, 46.4 and 23.9% samples were found positive. They reported presence of MAP in holy river Yamuna, this may be due presence of large number of goshalas (cow shelters) for religious reasons in the holy city of Vrindavan (where samples were collected) and most of these cow shelters wash off 'cow dung' into the drains leading to river Yamuna, rather than lifting cow dung making compost. However, water samples from river Chambal with little human and animal population on its banks was comparatively free of MAP and other pathogenic micro-organisms. Similarly soil samples from farm area, where goatherds are reared over the years had soil samples positive MAP as compared to farmer's fields. besides heavy load of other microbial contaminants in soil, water and environment.

### Risk associated with presence of MAP in human food chain

JD caused by MAP in animals is one of the most difficult diseases to diagnose and control because it has ability to resist adverse conditions in the natural environment. MAP is intracellular bacilli and JD infected animals, macrophages infected / laden with MAP can be found throughout the body of the animals. Most tissues including lymph nodes, spleen, bone

marrow, liver, kidneys, testis, fetuses, supra-mammary lymph node and lungs are affected (Kopecka et al., 2008). It has been suggested that meat harvested from old dairy cows and used for making ground beef for human consumption may be a active source of MAP infection (Rossiter et al., 2001). Meat can also be contaminated with fecal material loaded with MAP bacilli during slaughter and procedures used for evisceration and processing of meat.

JD infected animals shed MAP bacilli in abundant quantities onto the land and pastures and increase the load of bacilli in the environment. Unlike *Mycobacterium bovis*, MAP is able to survive in the environment (Rowe et al., 2006). The contact time (CT) values for the effect of chlorine on MAP have been estimated to be up to 580-2300 times greater than those for *E. coli* (Wiley et al., 2012). Information regarding other zoonotic pathogens surviving in the environment suggests that there is a high risk that MAP may spread through aerosols and drinking-water supplies (Herman-Taylor et al., 2000). MAP is present in pasteurized milk (Slana et al., 2008), infant formulation made from pasteurized milk (Lund et al., 2002; Makharia and Singh, 2009), breast milk of a woman suffering from CD (2009), cow manure “lagoons” that can leach into surface ground water, cow manure (solid and liquid forms) applied as fertilizer to agricultural land, and municipal tap water (Whiley et al., 2012) thus providing multiple routes of transmission to human population. European Commission DG - Health and Consumer Protection in 2000 commissioned a report from the Scientific Committee on Animal Health as well as Animal Welfare on the possible links between Crohn’s disease and MAP (EC, 2000). The National Association for Colitis and Crohn’s disease (NACC) in UK commissioned a report from an expert review group into the evidence linking MAP and Crohn’s disease in 2003 (NACC 2003). This report concluded that: ‘both live and dead MAP’ were present in human foods samples and the strongest evidence for this is in milk and DNA of MAP can be found in the bowel tissue of a proportion of patients with CD but also in lesser quantities in the bowel tissue of human beings having no lesions of CD. International Life Science Institute (ILSI), Europe in 2004 published a report from its Emerging Pathogen Task Force on MAP and the food chain (Gould et al., 2005). In its conclusion, it stated that “survival of MAP having public health concern depends on their possible involvement in human

disease, in particular CD. At the present time, despite substantial research, the possible involvement of MAP in human diseases is still under discussion. This opinion followed a review of MAP survival characteristics in food which noted in particular, studies demonstrating survival of viable MAP in pasteurized milk. Food Standards Australia New Zealand conducted a microbiological review of the association between JD and CD (FSANZ, 2004). It concluded that CD may be a “multi-factoral disease or syndrome, with no etiological factor appearing to dominate. At present there is inadequate scientific evidence to confirm a conclusive link between JD (MAP infection) in ruminants and some cases of CD in human beings”. The American Academy of Microbiology published a review on the evidence of pathogenicity in MAP (Nacy and Buckley, 2008). It has been noted that “there is a suspicion, supported by reports of genetic incompetence to interact correctly with certain bacteria and their products in some patients, that CD may have a currently unrecognized infectious origin, possibly environmentally derived”. It has also been noted that “the option of more than one infectious reason that leads to a similar set of indications confounds the research agenda to find both a cause and a cure for CD”. The report lists having five reasons why MAP has a suspected role in CD including MAP’s ability to survive milk pasteurization and the success in some CD patients of antibiotic therapy against *Mycobacteria*. The report lays out the pros and cons of a possible association between MAP and CD. Finally, they concluded that “more research support and substantial additional research effort by both scientists and clinicians are necessary before we will know whether CD has an infectious aetiology and whether MAP is the offender”.

There is lack of awareness in developing and under-developed countries on the presence and level of MAP infection in animals and human population (Singh et al., 2014a; 2014b). MAP has also been associated with various other human diseases / syndromes besides Crohn’s disease in human population. Information on the association of MAP with different human health problems is yet to be recognized and taken seriously by the medical world globally.

JD or MAP infection is primarily a disease of food-producing domestic livestock (cattle, buffaloes, sheep and goats). Food products (meat and milk) derived from animals infected with MAP is commonly

contaminated much before harvesting, that is during the disseminative stage of JD, when pathogen exist in muscle meat, internal organs (intestines, mesenteric lymph nodes), colostrum and milk, and the un-born fetus (Brady et al., 2008). During harvesting of milk and meat from animals, there is second opportunity for the contamination of food products with MAP by contact with faeces of the animal infected with JD (Grant, 2010). Thus raw products obtained from MAP-infected animals are likely to be contaminated with MAP (Eltholth et al., 2009). Counts up to 560 MAP/ml of raw milk from individual cows have been reported (Grant, 2010). The question then becomes whether manufacturing processes or, cooking by the consumer will reliably kill MAP.

During food manufacturing processes, killing of MAP is based on pasteurization, specifically high temperature short-time (HTST) pasteurization (Grant et al., 2002b). Early laboratory studies were carried out with MAP inoculated milk in test tubes. In these studies, the milk was subjected to heat treatments similar to those used for pasteurization in holding tanks (holder pasteurizing) or for high-temperature short-time (HTST) pasteurization (Stabel et al., 2004). MAP was recovered from most of the samples of pasteurized milk. Studies with milk heated in apparatus (test tubes, capillary tubes, or laboratory pasteurizers) indicated that both holder and HTST pasteurization could be effective for killing (inactivating) of MAP in milk. In measurement of standard thermal tolerance parameters (STTP) for MAP in laboratory studies indicate that it is more heat-resistant than *Mycobacterium bovis* and *Coxiella burnetti* (milk-borne zoonotic pathogens) designed to be killed by pasteurization (Sung & Collins, 1998). Corroborating this are 3 independent retail milk surveys reporting recovery of viable MAP by culture from retail HTST pasteurized milk (Ellingson et al., 2005). Clearly, HTST pasteurization method kills large numbers of MAP bacilli in milk. However, evidences suggests that it does not kill 100% of MAP bacilli and post pasteurization MAP contamination is frequently occurring. This may be happening due to formation of clumps by the bacilli due to high lipid contents during initial heating phase. Within these clumps these bacilli survive the pasteurization temperature.

#### **a. MAP in dairy products**

Majority of MAP based infections occur through food producing domestic livestock. Milk derived from

infected animals is generally contaminated before harvesting (milking). During harvesting of milk, there is a huge scope of contamination of milk and meat with fecal matter (Rademaker et al., 2007). Early studies were directed at investigating the presence of MAP in live and dead condition in raw and pasteurized milk using PCR technology (Ellingson et al., 2005). Irene Grant has extensively studied and published results in 2002: MAP presence was confirmed by culture in 1.6 and 1.8% of raw and pasteurized milk, respectively. Grant et al., (2002a) concluded that live MAP is infrequently present in commercially pasteurized milk in the UK. Ministry of Agriculture, fisheries and Food (MAFF), UK has started research that was continued by Food Standards agency, reported 1.7% prevalence of live MAP from pasteurized milk on retail sale (FSA, 2000) and pasteurized cow's milk was screened by IS900 PCR. Field studies were done on retail pasteurized cow's milk for screening of MAP infection by IS900 PCR in UK, though it was unable to distinguish between live and dead bacilli but study indicated high risk of transmission of MAP to human food chain (Miller et al. 1996). Queen's University Belfast screened PCR positive cow's milk samples with optimized decontamination protocols and immunomagnetic capture method where, 11.8% were positive, and by culture 1.8% of samples were positive for MAP (Grant et al. 2002a). In Switzerland, 19.7% of 1384 samples of bulk-tank milk were positive by IS900 PCR again emphasized the risk of MAP in human food chain (Corti & Stephan 2002). Bulk-tank milk were positive by IS900 PCR.

MAP is not inactivated during pasteurization; therefore live MAP bacilli are present in milk and milk products made from pasteurized milk. Extensive use of milk products made from pasteurized milk globally, poses serious threat to human life. Initially this problem was restricted to most of the developed countries due to high dependence on pasteurized milk and milk products. But slowly this problem has started plaguing developing countries like India due to increasing use of milk products like ice-cream, milk coffee and condensed milk made from pasteurized milk. Since the use of milk products made from pasteurized milk increased dramatically in the country, therefore chances of contamination of human population with MAP has also increased. Bio-load of MAP has been estimated in different types of milk and milk products made from pasteurized milk of domestic

livestock species in different parts of world as well as in India.

### **b. MAP in meat products**

MAP primarily affects the intestines and is found systemically in animal body with the advancing stages of JD (Rubery, 2002). In addition to the gastrointestinal system, MAP has also been detected in milk, blood, semen, mammary glands, lymph nodes, reproductive organs and fetuses (Larsen et al., 1970; Sweeney et al., 1992b; Kopecna *et. al.*, 2008). MAP can be detected in these tissues of animals with sub-clinical infection. Therefore, it is possible that meat can be contaminated with MAP obtained from both clinically infected and apparently healthy animals. Meat could also be become contaminated with faecal material during slaughtering and processing procedures (Verma et al., 2014).

In a study MAP has been detected in the intestinal lymph nodes / faeces of 34.0% of healthy thin dairy cows and from 3.0% of healthy thin beef cows at slaughter (Rossiter & Henning 2001). MAP was also isolated from 3.0% in peripheral lymph nodes and 8.0% in liver of dairy cows and beef cows, respectively. Information is not available concerning the ability of MAP to survive during meat cooking and processing methods. It is likely that contaminated meat that is cooked at low temperatures or not processed thoroughly may contain live MAP. Though, the significance of meat as a potential vehicle of food-borne exposure to MAP relative to the potential exposure through milk is not known (Food Standards Australia New Zealand, 2004). It has been reported that fecal matter of slaughtered animals is the main source of MAP contamination in meat and meat product (Wells et al., 2009). Gwozdz et al (2000) has reported the evidence of MAP DNA in the blood and / or liver of many sheep with advanced symptoms of JD.

### **Time period wise bio-load of MAP in domestic livestock species in last 30 years (1985–2015): CIRG study**

Of the 24,944 clinical samples (faecal, serum, blood) from goats, sheep, cattle and buffaloes screened by multiple diagnostic tests (faecal microscopy, faecal culture, serum ELISA and blood PCR) over the last 30 years (1985–2015), cumulative average bio-load of MAP was 25.5%. Livestock species-wise average bio-

load in goats, sheep, cattle and buffaloes was 21.5, 35.6, 43.5 and 28.6%, respectively. Bio-load was significantly lower in goats ( $P < 0.05$ ) than in sheep, buffaloes and cows. Average bio-load of MAP in 30 years (1985–2015) time period at five yearly intervals; 1985–90, 1991–95, 1996–2000, 2001–05, 2006–10 and 2011–15 was 11.4, 13.1, 11.1, 24.2, 28.9 and 42.8%, respectively and showed increasing trend. Time zone-wise, the average bio-load was 11.6, 27.0 and 38.8%, in the time period of 15 years (1985–2000), 10 years (2001–10) and 5 years (2011–15), respectively.

### **Time period-wise bio-load of MAP in human population in last 8 years (2008–2015): CIRG study**

Bio-load of MAP was estimated in human population from different region of the country by 'mass screening'. Of the 24,322 serum samples screened by Indigenous ELISA, 34.1% populations were positive for MAP infection. Percent bio-load of MAP infection was 34.0, 49.2 and 31.7% in Period A (2008-13), Period B (2013-14) and Period C (2014-15), respectively by Indigenous ELISA. Of 3, 224 blood samples screened by PCR, 8.7% population was MAP positive. Time period-wise bio-load of MAP infection was 8.4, 18.1 and 17.4%, respectively by blood PCR. Screening of 157 stool samples by stool PCR, the bio-load was 21.0% and percent bio-load of MAP was 5.9, 55.5 and 44.7 in Period A, Period B and Period C, respectively. Time period wise cumulative bio-load was 30.2, 47.8, 30.5 in period A, Period B and Period C, respectively.

### **Conclusions**

MAP is a very tough bacteria having strong zoonotic potential competent to infect wide range of hosts. It contaminates raw milk and meat derived from the infected animals and water runoff from farms contaminated with fecal matter of animals infected with MAP. Use of pasteurization and low heating conditions (roasting, smoking etc.) may help MAP to survive cooking and food manufacturing processes. There are lots of evidences suggesting, MAP is a potential candidate for causation of Crohn's disease. Meat, milk and other food products from animals infected with MAP serve as important means of transmission of MAP to human population through animal based food chain. The evidence are huge which

appear to indicate that MAP is proficient in surviving during food processing we applied to protect us from diseases, such as cooking and pasteurization. Clinical and pathological similarities of Crohn's disease with other mycobacterial diseases. Associated evidences suggest that presence of MAP in the environment and milk and meat supplies (animal-based food) are not safe and may help to spread MAP to human beings. Human infection can only be restricted, if domestic livestock population is cured of infection.

### Acknowledgments

The authors are highly thankful to Director, CIRG; ICMR project, ICMR, New Delhi and OPZD project, ICAR, New Delhi for strengthening facilities for research at CIRG, Makhdoom.

### Conflict of interests

The authors declare that they have no competing interests (conflict of interests).

### Funding

Indian Council of Medical Research [grant number 5/8/5/28/TF/2013/ECD-I].

### References

1. Aboagye G, Rowe MT. Occurrence of *Mycobacterium avium* subspecies *paratuberculosis* in raw water and water treatment operations for the production of potable water. *Wat. Res.* 2011; 45: 3271–3278.
2. Abubakar I, Myhill D, Aliyu SH, Hunter PR. Detection of *Mycobacterium avium* subspecies *paratuberculosis* from patients with Crohn's disease using nucleic acid-based techniques: a systematic review and meta-analysis. *Inflamm Bowel Dis.* 2008; 14: 401–410.
3. Aggarwal A, Chag M, Sinha N, Naik S. Takayasu's arteritis: role of *Mycobacterium tuberculosis* and its 65 kDa heat shock protein. *Int J Cardiol.* 1996; 55(1):49–55. [10.1016/0167-5273\(96\)02660-5](https://doi.org/10.1016/0167-5273(96)02660-5)
4. Ayele WY, Svastova P, Roubal P, Bartos M and Pavlik I. *Mycobacterium avium* subspecies *paratuberculosis* cultured from locally and commercially pasteurized cow's milk in the Czech republic. *Applied and environmental microbiology.* 2005; 71(3): 1210-1214.
5. Bannantine JP, Li L, Mwangi M, Cote R, Garay JAR, Kapur V. Complete Genome Sequence of *Mycobacterium avium* subsp. *paratuberculosis*, Isolated from Human Breast Milk. *Genome Announc.* 2014; 2 (1): e01252-13.
6. Bauman CA, Jones-Bitton A, Menzies P, Toft N, Jansen J, Kelton D. Prevalence of paratuberculosis in the dairy goat and dairy sheep industries in Ontario, Canada. *Can Vet J.* 2016; 57(2): 169–175.
7. Beumer A, King D, Donohue M, Mistry J, Covert T, Pfaller S. Detection of *Mycobacterium avium* subspecies *paratuberculosis* in drinking water and biofilms by quantitative pcr. *Appl. Environ. Microbiol.* 2010; 76: 7367–7370.
8. Blanchard DC, Syzdek LD. Water-to-Air Transfer and Enrichment of Bacteria in Drops from Bursting Bubbles. *Applied and environmental microbiology.* 1982;43:1001–1005.
9. Blau B. Familial granulomatous arthritis, iritis, and rash. *J Pediatr.* 1985; 107(5):689–93. [10.1016/S0022-3476\(85\)80394-2](https://doi.org/10.1016/S0022-3476(85)80394-2)
10. Botsaris G, Slana I, Liapi M, Dodd C, Economides C, Rees C, Pavlik I. Rapid detection methods for viable *Mycobacterium avium* subspecies *paratuberculosis* in milk and cheese. *International Journal of Food Microbiology.* 2013; 141:S87-S90
11. Brady C, O'Grady D, O'Meara F, Egan J, Bassett H. Relationships between clinical signs, pathological changes and tissue distribution of *Mycobacterium avium* subspecies *paratuberculosis* in 21 cows from herds affected by JD. *Vet Rec.* 2008; 162 (5): 147-52.
12. Carvalho IA, Pietralonga PAG, Schwarz DGG, Faria ACS, Moreira MAS. Short Communication: Recovery of viable *Mycobacterium avium* subspecies *paratuberculosis* (MAP) from retail pasteurized whole milk in Brazil. *Journal of Dairy Science.* 2012; 95 (12):6946-8. [Doi:10.3168/jds.2012-5657](https://doi.org/10.3168/jds.2012-5657).
13. Chamberlin W, Borody T, Naser S. MAP-associated Crohn's disease MAP, Koch's postulates, causality and Crohn's disease. *Dig Liver Dis.* 2007; 39: 792–794.
14. Chiodini RJ, Hermon-Taylor J. The thermal resistance of *Mycobacterium paratuberculosis* in raw milk under conditions simulating pasteurization. *J. Vet. Diagn. Invest.* 1993; 5:629–631.

15. Chiodini RJ, Van Kruiningen HJ, Merkal RS. Ruminant paratuberculosis (Johne's disease): the current status and future prospects. *Cornell Vet.* 1984; 74: 217-262.
16. Cirillo JD, Falkow S, Tompkins LS, Bermudez LE. Interaction of *Mycobacterium avium* with environmental amoebae enhances virulence. *Infect Immun.* 1997; 65, 3759–3767.
17. Clark DL Jr, Anderson JL, Koziczkowski JJ, Elligson JLE. Detection of *Mycobacterium avium* subspecies *paratuberculosis* genetic components in retail cheese curds purchased in Wisconsin and Minnesota by PCR. *Mol. Cell. Probes.* 2006; 20: 197-202.
18. Collins MT, Miliotis MD, Bier JW. *International Handbook of Foodborne Pathogens.* Boca Raton, FL: CRC Press; 2003. 17 p.
19. Collins MT, Spahr U, Murphy PM. Ecological characteristics of *M. paratuberculosis*, *In Bulletin of the International Dairy Federation*, no. 362/2001. International Dairy Federation, Brussels, Belgium. 2001. p. 32-40.
20. Collins MT. *Mycobacterium paratuberculosis*: a potential food-borne pathogen. *J Dairy Sci.* 1997; 80: 3445-3448.
21. Collins MT. Paratuberculosis: review of present knowledge. *Acta Vet Scand.* 2003; 44:217–21.
22. Corti, S. and R. Stephan. Detection of *Mycobacterium avium* subspecies *paratuberculosis* specific IS900 insertion sequences in bulk-tank milk samples obtained from different regions throughout Switzerland. *BMC Microbiol.* 2002; 2:15.
23. Cossu D, Cocco E, Paccagnini D, Masala S, Ahmed N, Frau J, et al. Association of *Mycobacterium avium* subsp. *paratuberculosis* with multiple sclerosis in Sardinian patients. *PLoS One.* 2011; 6(4):e18482.10.1371/journal.pone.0018482
24. Dalziel TK. Chronic interstitial enteritis. *Dis Colon Rectum.* 1989; 32:1076–1078.
25. Danieli MG, Candela M, Ricciatti AM, Reginelli R, Danieli G, Cohen IR, et al. Antibodies to mycobacterial 65 kDa heat shock protein in systemic sclerosis (scleroderma). *J Autoimmun.* 1992; 5(4):443–5.10.1016/0896-8411(92)90004-A
26. De Hertogh G, Aerssens J, Geboes KP, Geboes K. Evidence for the involvement of infectious agents in the pathogenesis of Crohn's disease. *World J Gastroenterol.* 2008; 14: 845–852.
27. Desio D, Nizza S, Montagnaro S, Sasso S, De Martino L, Iovane V, et al. Estimated Prevalence of Johne's Disease in Herds of Water Buffaloes (*Bubalus Bubalis*) in the Province of Caserta. *Italian journal of animal science.* 2013; 12:1-9.
28. Dhand NK, Eppleston J, Whittington RJ, Toribio J-ALML. Association of farm soil characteristics with ovine Johne's disease in Australia. *Prev. Vet. Med.* 2009; 89:110–120.
29. Direskeneli H, Saruhan-Direskeneli G. The role of heat shock proteins in Behcet's disease. *Clin Exp Rheumatol.* 2003; 21(30):S44–8.
30. Dow CT, Ellingson JL. Detection of *Mycobacterium avium* ss. Paratuberculosis in Blau syndrome tissues. *Autoimmune Dis.* 2010; 20(2010):127692.10.4061/2010/127692
31. Dow CT. *Mycobacterium paratuberculosis* and autism: is this a trigger?. *Med Hypotheses.* 2011; 77(6):977-81. doi: 10.1016/j.mehy.2011.08.024.
32. Ellingson JLE, Anderson JL, Koziczkowski JJ, et al. Detection of viable *Mycobacterium avium* subsp. *paratuberculosis* in retail pasteurized whole milk by two culture methods and PCR. *J Food Prot.* 2005; 67; 966-72.
33. Eltholth MM, Marsh VR, Van Winden S, Guitian FJ. Contamination of food products with *Mycobacterium avium* subspecies *paratuberculosis*: a systemic review. *J Appl Microbiol* 2009; 107: 1061-71.
34. European Commission. European commission report. Possible links between Crohn's disease and paratuberculosis. Report of the Scientific Committee on Animal Health and Animal Welfare. 2000. Report No: SANCO/B3/R16/2000. Available on line at: [http://europa.eu.int/comm/food/fs/sc/scah/out38\\_en.pdf](http://europa.eu.int/comm/food/fs/sc/scah/out38_en.pdf)



35. Falkingham JO. Ecology of nontuberculous mycobacteria-where do human infections come from? Preface. *Sem. Resp. Crit. Care.* 2013; 34: 95–102.
36. Feazel LM, Baumgartner LK, Peterson KL, Frank DN, Harris JK, Pace NR. Opportunistic pathogens enriched in showerhead biofilms. *P. Natl. Acad. Sci. USA.* 2009; 106:16393–16398.
37. Feller M, Huwiler K, Stephan R, Altpeter E, Shang A, Furrer H, et al. *Mycobacterium avium* subspecies *paratuberculosis* and Crohn's disease: a systematic review and meta-analysis. *Lancet Infect Dis.* 2007; 7: 607–613.
38. Food Standards Australia New Zealand. Association between Johne's disease and Crohn's disease. A microbiological review. Technical report series No. 35.2004. p 1-21. <http://www.foodstandards.gov.au>.
39. Gao A, Mutharia L, Chen S, Rahn K, Odumeru J. Effect of pasteurization on survival of *Mycobacterium paratuberculosis* in milk. *J. Dairy Sci.* 2002; 85:3198–3205.
40. Gill, C.O.; Saucier, L.; Meadus, W.J. *Mycobacterium avium* subsp *paratuberculosis* in dairy products, meat, and drinking water. *J. Fd Prot.* 2011; 74: 480–499.
41. Gould G, Franken P, Hammer P, Mackey B, Shanahan F. *Mycobacterium avium* ssp. *paratuberculosis* and the food chain. *Food Prot. Trends.* 2005; 25:268–297.
42. Grant IR, Ball HJ, Neill SD, Rowe MT. Inactivation of *Mycobacterium paratuberculosis* in cows' milk at pasteurization temperatures. *Appl. Environ. Microbiol.* 1996. 62:631–636.
43. Grant IR, Ball HJ, Rowe MT. Effect of high temperature, short-time (HTST) pasteurization on milk containing low numbers of *Mycobacterium paratuberculosis*. *Lett. Appl. Microbiol.* 1998; 26:166–170.
44. Grant IR, Ball HJ, Rowe MT. Effect of higher pasteurization temperatures, and longer holding times at 72°C, on the inactivation of *Mycobacterium paratuberculosis* in milk. *Lett. Appl. Microbiol.* 1999; 28:461–465.
45. Grant IR, Ball HJ, Rowe MT. Incidence of *Mycobacterium avium* subspecies *paratuberculosis* in bulk raw and commercially pasteurized cows' milk from approved dairy processing establishment in the united kingdom. *Appl Env Microbiol* 2002a; 68:428-35.
46. Grant IR, Hitchings EI, McCartney A, Ferguson F, Rowe MT. Effect of commercial-scale high-temperature, short-time pasteurization on the viability of *Mycobacterium avium* subspecies *paratuberculosis* in naturally infected cows' milk. *Appl Env Microbiol.* 2002b; 68 (2): 602-7.
47. Grant IR. *Mycobacterium avium* subspecies *paratuberculosis* in animal derived foods and the environment. In: Behr MA, Collins DM, editors. *Paratuberculosis: organism, disease, control.* Oxfordshire, UK: CABI; 2010. P. 29-39.
48. Grant IR. Zoonotic potential of *Mycobacterium avium* ssp. *paratuberculosis*: the current position. *Journal of Applied Microbiology.* 2005; 98: 1282-1293.
49. Greenstein RJ. Is Crohn's disease caused by a mycobacterium? Comparisons with leprosy, tuberculosis, and Johne's disease. *Lancet Infect Dis.* 2003; 3: 507–514.
50. Grewal SK, Rajeev S, Sreevatsan S, Michel FC Jr. Persistence of *Mycobacterium avium* subsp. *paratuberculosis* and other zoonotic pathogens during simulated composting, manure packing, and liquid storage of dairy manure. *Appl Environ Microbiol.* 2006.72: 565–574.
51. Gwozdz JM, Thompson KG, Murray A, West DM, Manktelow BW. Use of the polymerase chain reaction assay for the detection of *Mycobacterium avium* subspecies *paratuberculosis* in blood and liver biopsies from experimentally infected sheep. 2000; 78(9):622–624. DOI: 10.1111/j.1751-0813.2000.tb11938.x
52. Haghkhah M, Ansari-Lari M, Novin-Baheran AM, Ayatollah B. Herd-level prevalence of *Mycobacterium avium* subspecies *paratuberculosis* by bulk-tank milk PCR in Fars province (Southern Iran) dairy herds. *Prev Vet Med.* 2008; 86:8-13.
53. Hendrick S, Duffield T, Leslie K, Lissemore K, Archambault M, Kelton D. The prevalence of milk and serum antibodies to *Mycobacterium avium* subspecies *paratuberculosis* in dairy herds in Ontario. *Canadian Veterinary Journal-Revue Veterinaire Canadienne.* 2005a; 46:1126–1129.
54. Hendrick SH, Duffield TF, Kelton DF, Leslie KE, Lissemore KD, Archambault M. Evaluation of enzyme-linked immunosorbent assays performed on milk and serum samples for detection of

- paratuberculosis in lactating dairy cows. *Journal of the American Veterinary Medical Association*. 2005b;226: 424–428.
55. Hendrick SH, Duffield TF, Leslie KE, Lissemore KD, Archambault M, Bagg R, et al. Monensin might protect Ontario, Canada dairy cows from paratuberculosis milk-ELISA positivity. *Preventive Veterinary Medicine*. 2006; 76: 237–248.
  56. Hermon-Taylor J, Barnes N, Clarke C, Finlayson C. *Mycobacterium paratuberculosis* cervical lymphadenitis, followed five years later by terminal ileitis similar to Crohn's disease. *BMJ*. 1998. 316: 449–453.
  57. Hermon-Taylor J, Bull TJ, Sheridan JM, Cheng J, Stellakis ML, Sumar N. Causation of Crohn's disease by *Mycobacterium avium* subspecies *paratuberculosis*. *Can. J. Gastroenterol*. 2000. 14:521–539.
  58. Hermon-Taylor, J. Causation of Crohn's disease: the impact of clusters. *Gastroenterology*. 1993.104:643-646.
  59. Hruska K, Bartos M, Kralik P, Pavlik I. *Mycobacterium avium* subsp. *paratuberculosis* in powdered infant milk: Paratuberculosis in cattle—the public health problem to be solved. *Veterinarni Medicina*. 2005; 50: 327–335.
  60. Hruska K, Kaevska M. Mycobacteria in water, soil, plants and air: A review. *Vete. Med*. 2012; 57, 623–679. 27.
  61. Hruska K, Slana I, Kralik P, Pavlik I. *Mycobacterium avium* subsp. *Paratuberculosis* in powdered infant milk: F57 competitive real time PCR. *Veterinarni Medicina*. 2011; 56: 226-230.
  62. Hussain MH, Saqib M, Al-Maawali MG, Al-Makhladi S, Al-Zadjali MS, Al-Sidairi T, et al. Seroprevalence of *Mycobacterium avium* subspecies *paratuberculosis* (MAP) and evaluation of risk factors in camels of the Sultanate of Oman. *Trop Anim Health Prod*. 2015; 47:383-389.
  63. Ikonopoulou J, Pavlik I, Bartos M, Svastova P, Ayele WY, Roubal P, et al. Detection of *Mycobacterium avium* subsp. *paratuberculosis* in retail cheeses from Greece and the Czech Republic. *Appl. Environ. Microbiol*. 2005; 71: 8934–8936.
  64. Jaravata CV, Smith WL, Rensen GJ, Ruzante J, Cullor JS. “Survey of Ground Beef for the Detection of *Mycobacterium avium paratuberculosis*.” *Foodborne Pathogens and Disease*. 2007; 4(1): 103-106.
  65. Jayarao BM, Pillai SR, Wolfgang DR, Griswold DR, Rossiter CA, Tewari D, Burns CM, Hutchinson LJ. Evaluation of IS900-PCR assay for detection of *Mycobacterium avium* subspecies *paratuberculosis* infection in cattle using quarter milk and bulk tank milk samples. *Foodborne Pathogens and Disease*. 2004; 1, 17–26.
  66. Jeyanathan M, Boutros-Tadros O, Radhi J, Semret M, Bitton A, Behr MA. Visualization of *Mycobacterium avium* in Crohn's tissue by oil-immersion microscopy. *Microbes Infect*. 2007; 9: 1567–1573.
  67. Jones PH, Farver TB, Beaman B, Cetinkaya B, Morgan KL. Crohn's disease in people exposed to clinical cases of bovine paratuberculosis. 2006. *Epidemiol Infect*. 2006;134:49-56.
  68. Kaevska M, Lvoncik S, Lamka J, Pavlik I, Slana I. Spread of *Mycobacterium avium* subsp. *paratuberculosis* through soil and grass on a Mouflon (*Ovis aries*) pasture. *Curr Microbiol*. 2014; 69(4):495–500.10.1007/s00284-014-0618-418
  69. Kaur P, Filia G, Singh SV, Patil PK, Sandhu KS. Molecular detection and typing of *Mycobacterium avium* subspecies *paratuberculosis* from milk samples of dairy animals. *Trop. Anim. Health Pro*. 2010; 42: 1031- 1035.
  70. Kawasaki E. Type 1 diabetes and autoimmunity. *Clin Pediatr Endocrinol*. 2014; 23(4):99–105.10.12977/cpe.23.99
  71. Keswani J, Frank JF. Thermal inactivation of *Mycobacterium paratuberculosis* in milk. *J. Food Prot*. 1998; 61:974–978.
  72. Klanicova B, Seda J, Slana I, Slany M, Pavlik I. The tracing of mycobacteria in drinking water supply systems by culture, conventional, and real time pcrs. *Curr. Microbiol*. 2013; 67:725–731.
  73. Klausen J, Huda A, Ekeroth L, Ahrens P. Evaluation of serum and milk ELISAs for paratuberculosis in Danish dairy cattle. *Preventive Veterinary Medicine*. 2003; 58:171–178.

74. Kopečna M, Parmová I, Dvorská-Bartosová L, Moravková M, Babák V, Pavlík I. Distribution and transmission of *Mycobacterium avium* subspecies *paratuberculosis* in farmed red deer (*Cervus elaphus*) studied by faecal culture, serology and IS900 RFLP examinations. *Vet. Med.* 2008; 53:510–523
75. Kruze J, Monti G, Schulze F, Mella A, Leiva S. Herd-level prevalence of Map infection in dairy herds of southern Chile determined by culture of environmental fecal samples and bulk-tank milk qPCR, *Preventive Veterinary Medicine.* 2013. 111; 319–324.
76. KuKanich KS, Vinasco J, Scott HM. Detection of *Mycobacterium avium* subspecies *paratuberculosis* from intestinal and nodal tissue of dogs and cats. *ISRN Vet Sci.* 2013;2013:1-4.
77. Kumar S, Singh SV, Sevilla I, Singh AV, Whittington RJ, Juste RA, et al. Lacto–incidence and evaluation of 3 tests for the diagnosis of Johne’s disease using milk of naturally infected goatherds and genotyping of *Mycobacterium avium* subspecies *paratuberculosis*. *Small Rumin. Res.* 2008a; 74: 37.
78. Kumar S, Singh SV, Singh AV, Singh PK, Sohal JS, Maitra A. Wildlife (*Boselaphus tragocamelus*)-small ruminant (goat and sheep) interface in the transmission of ‘Bison type’ genotype of *Mycobacterium avium* subspecies *paratuberculosis* in India. *Comp Immunol Microbiol Infect Dis.* 2010; 33: 145-59.
79. Larsen AB, Kopecky KE. *Mycobacterium paratuberculosis* in reproductive organs and semen of bulls. *Am J Vet Res.* 1970; 31: 255-259.
80. Lombard JE, Byrem TM, Wagner BA, McCluskey BJ. Comparison of milk and serum enzyme-linked immunosorbent assays for diagnosis of *Mycobacterium avium* subspecies *paratuberculosis* infection in dairy cattle. *Journal of Veterinary Diagnostic Investigation.* 2006a; 18, 448–458.
81. Lombard JE, Wagner BA, Smith RL, McCluskey BJ, Harris BN, Payeur JB, et al. Evaluation of environmental sampling and culture to determine *Mycobacterium avium* subspecies *paratuberculosis* distribution and herd infection status on US dairy operations. *Journal of Dairy Science.* 2006b; 89: 4163–4171.
82. Lorencová A, Vasicková P, Makovcová J, Slaná I. Presence of *Mycobacterium avium* subspecies and hepatitis E virus in raw meat products. *J Food Prot.* 2014; 77(2):335-8. doi: 10.4315/0362-028X.JFP-13-252.
83. Lund BM, Gould GW, Rampling AM. Pasteurization of milk and the heat resistance of *Mycobacterium avium* subsp. *paratuberculosis*: a critical review of the data. *Int J Food Microbiol* 2002;77:135–45. 39.
84. Mayberry JF, Hitchens RA. Distribution of Crohn’s disease in Cardiff. *Soc Sci Med.* 1978;12:137–138. doi: 10.1016/S0277-9536(78)80021-5.
85. MCNeese AL; Markesich D, Zayyani NR, Graham DY. *Mycobacterium paratuberculosis* as a cause of Crohn’s disease. *Expert review of Gastroenterology & Hepatology.* 2015; 9 (12): 1523-34.
86. Miller D, Ford D, Sanderson J, Withey S, Tizard M, Doran T, et al. IS900 PCR to detect *Mycobacterium paratuberculosis* in retail supplies of whole pasteurised cows’ milk in England and Wales. *Applied and Environmental Microbiology.* 1996; 62:3446–3452.
87. Miyata M, Kogure A, Sato H, Kodama E, Watanabe H, Ohira H, et al. Detection of antibodies to 65 KD heat shock protein and to human superoxide dismutase in autoimmune hepatitis-molecular mimicry between 65 KD heat shock protein and superoxide dismutase. *Clin Rheumatol.* 1995;14(6):673–7. doi:10.1007/BF02207935
88. Momotani E, Romona NM, Yoshihara K, Momotani Y, Hori M, Ozaki H, Eda S, Ikegami M. Molecular pathogenesis of bovine paratuberculosis and human inflammatory bowel diseases. *Veterinary Immunology and Immunopathology.* 2012;148: 55-68.
89. Moudgil KD, Chang TT, Eradat H, Chen AM, Gupta RS, Brahn E, et al. Diversification of T cell responses to carboxy-terminal determinants within the 65 kD heat-shock protein is involved in regulation of autoimmune arthritis. *J Exp Med.* 1997;185(7):1307–16. doi:10.1084/jem.185.7.1307

90. Muehlherr JE, Zweifel C, Corti S, Blanco JE, Stephan R. Microbiological quality of raw goat's and ewe's bulk-tank milk in Switzerland. *J. Dairy Sci.* 2003; 86:3849–3856.
91. Nacy C, Buckley M. *Mycobacterium avium paratuberculosis*: Infrequent human pathogen or public health threat? A report from the American Academy of Microbiology. 2008: 37 pp.
92. Naser SA, Collins MT, Crawford JT, Valentine JF. Culture of *Mycobacterium avium* subspecies *paratuberculosis* (MAP) from the Blood of Patients with Crohn's disease: A Follow-Up Blind Multi Center Investigation. *The Open Inflammation Journal.* 2009; 2: 22-23.
93. Naser SA, Ghobrial G, Romero C, Valentine JF. Culture of *Mycobacterium avium* subspecies *paratuberculosis* from the blood of patients with Crohn's disease. *Lancet.* 2004; 364: 1039–1044.
94. Naser SA, Schwartz D, Shafran I. Isolation of *Mycobacterium avium* subsp. *paratuberculosis* from breast milk of Crohn's disease patients. *Am J Gastroenterol.* 2000; 95: 1094-1095.
95. National Association for Colitis and Crohn's Disease. The report of the NACC expert review group into the evidence linking *Mycobacterium paratuberculosis* (Map) and Crohn's disease. 2003. Accessed at: <http://nacc.org.uk/MAPver9.pdf>
96. Nazareth N, Magro F, Machado E, Ribeiro TG, Martinho A, Rodrigues P, Alves R, Macedo GN, Gracio D, Coelho R, Abreu C, Appelberg R, Dias C, Macedo G, Bull T, Sarmiento A. Prevalence of *Mycobacterium avium* subsp. *Paratuberculosis* and *Escherichia coli* in blood samples from patients with inflammatory bowel disease. *Med Microbiol Immunol.* 2015; 204 (6): 681-692. doi: 10.1007/s00430-015-0420-3.
97. Nielsen SS, Ersboll AK. Age at occurrence of *Mycobacterium avium* subspecies *paratuberculosis* in naturally infected dairy cows. *Journal of Dairy Science.* 2006; 89:4557–4566.
98. Nielsen SS, Grohn YT, Enevoldsen C. Variation of the milk antibody response to paratuberculosis in naturally infected dairy cows. *Journal of Dairy Science.* 2002;85, 2795–2802.
99. Nielsen SS, Thamsborg SM, Houe H, Bitsch V. Bulk-tank milk ELISA antibodies for estimating the prevalence of paratuberculosis in Danish dairy herds. *Preventive Veterinary Medicine.* 2000; 44:1–7.
100. O'Reilly CE, O'Connor L, Anderson W, Harvey P, Grant IR, Donaghy J, et al. Surveillance of bulk raw and commercially pasteurized cows' milk from approved Irish liquid-milk pasteurization plants to determine the incidence of *Mycobacterium paratuberculosis*. *Appl. Environ. Microbiol.* 2004; 70:5138–5144.
101. Okara H, Nielsen SS, Toft N. Prevalence of *Mycobacterium avium* subsp. *paratuberculosis* infection in adult Danish non-dairy cattle sampled at slaughter. *Prev. Vet. Med.* 2010; 94: 185–190.
102. Paolicchi F, Cirone K, Morsella C, Gioffré A. First isolation of *Mycobacterium avium* subsp *Paratuberculosis* from commercial pasteurized milk in Argentina. *Braz J Microbiol.* 2012; 43(3): 1034–1037. doi: 10.1590/S1517-838220120003000028
103. Paolicchi FA, Zumarraga MJ, Gioffre A, Zamorano P, Morsella C, Verna A, Cataldi A, Alito A, Romano M. Application of different methods for the diagnosis of paratuberculosis in a dairy cattle herd in Argentina. *Journal of Veterinary Medicine Series Infectious Diseases and Veterinary Public Health.* 2003; 50:20–26.
104. Pavlík I, Mátlová L, Bártl J, Švastová P, Dvorská L, Whitlock R. Parallel faecal and organ *Mycobacterium avium* subsp. *paratuberculosis* culture of different productivity types of cattle. *Vet Microbiol.* 2000; 7: 7309–324.
105. Pedley S, Bartram J, Rees G, Dufour A, Cotruvo JAE. Pathogenic mycobacteria in water – A guide to public health consequences, monitoring and management. *Emerging Issues in Water and Infectious Disease Series, World Health Organization titles with IWA Publishing.* 2004; 1-222.
106. Pickup RW, Rhodes G, Arnott S, Sidi-Boumedine K, Bull TJ, Weightman A, et al. *Mycobacterium avium* subsp. *paratuberculosis* in the catchment area and water of the River Taff in South Wales, United Kingdom, and its potential relationship to clustering of Crohn's disease cases in the city of Cardiff. *Appl Environ Microbiol.* 2005; 71: 2130–2139.
107. Pickup RW, Rhodes G, Bull TJ, Arnott S, Sidi-Boumedine K, Hurley M, et al. *Mycobacterium avium* subsp. *paratuberculosis* in lake catchments, in

- river water abstracted for domestic use, and in effluent from domestic sewage treatment works: diverse opportunities for environmental cycling and human exposure. *Appl Environ Microbiol.* 2006; 72: 4067–4077.
108. Pillai SR, Jayarao BM. Application of IS900 PCR for detection of *M. avium* subsp. *paratuberculosis* directly from raw milk. *J Dairy Sci.* 2002; 85:1052-1057.
109. Pinna A, Masala S, Blasetti F, Maiore I, Cossu D, Paccagnini D, et al. Detection of serum antibodies cross-reacting with *Mycobacterium avium* subspecies *paratuberculosis* and beta-cell antigen zinc transporter 8 homologous peptides in patients with high-risk proliferative diabetic retinopathy. *PLoS One.* 2014; 9(9):e107802. doi:10.1371/journal.pone.0107802
110. Rademaker JLW, Vissers MMM, Te Giffel MC. Effective heat inactivation of *Mycobacterium avium* subsp. *paratuberculosis* in raw milk contaminated with naturally infected feces. *Appl. Environ. Microbiol.* 2007; 73:4185–4190.
111. Raghuvanshi T, Singh SV, Sharma RB, Gupta S, Chaubey KK, Kumar N, Dhama K. Identification of *Mycobacterium Avium* Subspecies *paratuberculosis* in Fresh Cheese (Paneer) from Goat Herds Endemic for Johne's Disease. *Journal of Infection and Molecular Biology.* 2013; 1: 46-48.
112. Raghuvanshi TS, Sharma RB, Singh AV, Singh B, Singh SV, Dhama K. 'Indigenous milk ELISA kit' vis a vis multiple test regime for the estimation of lacto-prevalence of *Mycobacterium avium* subspecies *paratuberculosis* in goatherds endemic for Johne's disease. *Indian J Comp. Microbiol. Immunol. Infect. Dis.* 2010; 31(1–2): 41–43.
113. Raizman EA, Habteselassie MY, Wu CC, Lin TL, Negron M, Turco RF. Leaching of *Mycobacterium avium* subsp *paratuberculosis* in soil under in vitro conditions. *Veterinary Medicine International.* 2011; 2011: 1-6. Article ID: 506239. doi:10.4061/2011/506239
114. Rawat KD, Chaudhary S, Kumar N, Gupta S, Chaubey KK, Singh SV, et al. Economic losses in a commercial dairy farm due to the outbreak of Johne's disease in India. *Res J Vet Practitioners.* 2014; 2:73-77.
115. Reid JD, Chiodini RJ. Serologic reactivity against *Mycobacterium paratuberculosis* antigens in patients with sarcoidosis. *Sarcoidosis.* 1993;10:32–35.
116. Rossiter CJ, Henning WR. Isolation of *Mycobacterium paratuberculosis* from thin market cows at slaughter. *J Anim Sci.* 2001. 79:113.
117. Rowan NJ, MacGregor SJ, Anderson JG, Cameron D, Farish O. Inactivation of *Mycobacterium paratuberculosis* by Pulsed Electric Fields. *Appl Environ Microbiol.* 2001; 67(6): 2833–2836. doi: 10.1128/AEM.67.6.2833-2836.2001
118. Rowe MT, Grant IR. *Mycobacterium avium* ssp. *paratuberculosis* and its potential survival tactics. *Lett Appl Microbiol.* 2006;42:305–311. doi: 10.1111/j.1472-765X.2006.01873.x.
119. Rubery E. A review of the evidence for a link between exposure to *Mycobacterium Paratuberculosis* (MAP) and Crohn's disease (CD) in humans. A report for the Food Standards Agency. Cambridge. 2002. 1-70.
120. Salgado M, Collins MT, Salazar F, Kruze J, Bölske G, Söderlund R, Juste R, Sevilla IA, Biet F, Troncoso F, Alfaro M. Fate of *Mycobacterium avium* subsp. *paratuberculosis* after application of contaminated dairy cattle manure to agricultural soils. *Applied and Environmental Microbiology.* 2011; 77:2122-2129.
121. Salgado M, Kruze J, Collins MT. Diagnosis of paratuberculosis by fecal culture and ELISA on milk and serum samples in two types of Chilean dairy goat herds. *Journal of Veterinary Diagnostic Investigation.* 2007; 19, 99–102.
122. Savi R, Ricchi M, Cammi G, Garbarino C, Leo S, Pongolini S, et al. Survey on the presence of *Mycobacterium avium* subsp. *paratuberculosis* in ground beef from an industrial meat plant. *Vet Microbiol.* 2015;177(3-4):403-8. doi: 10.1016/j.vetmic.2015.03.013.
123. Scanu AM, Bull TJ, Cannas S, Sanderson JD, Sechi LA, Dettori G, et al. *Mycobacterium avium* subspecies *paratuberculosis* infection in cases of irritable bowel syndrome and comparison with Crohn's disease and Johne's disease: common neural and immune pathogenicities. *J Clin Microbiol.* 2007; 45:3883–3890

124. Sechi LA, Scanu AM, Molicotti P, Cannas S, Mura M, Dettori G, et al. Detection and Isolation of *Mycobacterium avium* subspecies *paratuberculosis* from intestinal mucosal biopsies of patients with and without Crohn's disease in Sardinia. *Am J Gastroenterol.* 2005; 100: 1529–1536.
125. Shankar H, Singh SV, Singh PK, Singh AV, Sohal JS, Greenstein RJ. Presence, characterization, and genotype profiles of *Mycobacterium avium* subspecies *paratuberculosis* from unpasteurized individual and pooled milk, commercial pasteurized milk, and milk products in India by culture, PCR, and PCR–REA methods. *Int. J. Infect. Dis.* 2010; 14: e121–126.
126. Shankar H, Singh SV, Singh PK, Singh AV, Sohal JS. Recovery of live *Mycobacterium avium* subspecies *paratuberculosis* from milk and milk products from North India. *Indian J. Comp. Microbiol. Immunol. Inf. Dis.* 2008; 29: 31-35.
127. Sharma G, Singh SV, Sevilla I, Singh AV, Whittington RJ, Juste RA, Kumar S, Gupta VK, Singh PK, Sohal JS, Vihan VS. Evaluation of indigenous milk ELISA with m–culture and m–PCR for the diagnosis of bovine Johne's disease (BJD) in lactating Indian dairy cattle. *Res. Vet. Sci.* 2008; 84(1): 30–7.
128. Singh AV, Chauhan DS, Kumar A, Singh PK, Singh SV. Potential Etiologic Link and Association between *Mycobacterium avium* subspecies *paratuberculosis* and Crohn's Disease in Humans. *Research & Reviews: A Journal of Immunology.* 2012a; 2: 20-33.
129. Singh AV, Singh SV, Makharia GK, Singh PK, Sohal JS. Presence and characterization of *Mycobacterium avium* subspecies *paratuberculosis* from clinical and suspected cases of Crohn's disease and in the healthy human population in India. *Int J Infect Dis.* 2008; 12: 190-7.
130. Singh AV, Singh SV, Singh PK, Sohal JS, Singh MK. High prevalence of *Mycobacterium avium* subspecies *paratuberculosis* ('Indian bison type') in animal attendants suffering from gastrointestinal complaints who work with goat herds endemic for Johne's disease in India. *Int J Infect Dis.* 2011; 15: 677-683.
131. Singh SV, Kumar N, Sohal JS, Singh AV, Singh PK, Agarwal ND, Gupta S, Chaubey KK, Kumar A, Rawat KD, Deb R, Dhama K. First mass screening of the human population to estimate the bio-load of *Mycobacterium avium* subspecies *paratuberculosis* in North India. *J. Biol. Sci.* 2014b; 14(4): 237-247.
132. Singh SV, Singh AV, Singh R, Sandhu KS, Singh PK, Sohal JS, et al., Evaluation of highly sensitive indigenous milk ELISA kit with fecal culture, milk culture and fecal-PCR for the diagnosis of bovine Johne's disease (BJD) in India. *Comparative Immunol. Microbiol. Infect. Dis.* 2007a; 30: 175-186.
133. Singh SV, Singh AV, Singh R, Sandhu KS, Singh PK, Sohal JS. et al. Comparisons of multiple diagnostic tests for the diagnosis of bovine *paratuberculosis* in lactating cows. *Indian J. Anim. Sci.*, 2007b; 77: 1115-1117.
134. Singh SV, Singh PK, Singh AV, Sohal JS, Kumar N, Chaubey KK, et al. Bio-load and bio-type profiles of *Mycobacterium avium* subspecies *paratuberculosis* infection in the farm and farmer's herds / flocks of domestic livestock: A 28 years study (1985-2013). *Transboundary and Emerging Diseases.* 2014a; 61: 43–55.
135. Singh SV, Stephen BJ, Singh M, Gupta S, Chaubey KK, Sahzad, Jayaraman S, Aseri GK, Sohal JS, Bhatia AK, Pachoori A, Chauhan C and Dhama K. Evaluation of milk dot ELISA as 'Field Based Test' vis a vis milk plate ELISA for the detection of *Mycobacterium avium* subspecies *paratuberculosis* infection in lactating domestic livestock. *Indian Journal of Biotechnology.* 2016; 15:166-171.
136. Singh SV, Thakur S, Agarwal ND, Kumar N, Misri J, Gupta S, et al. Screening of human population for type 1 diabetes and *Mycobacterium avium* subspecies *paratuberculosis* from Chattarpur district of Madhya Pradesh (India). *Biomed Res Int.* 2014c; 2:612-619.
137. Singh SV, Tiwari AV, Singh PK, Singh B, Kumar A, Gururaj K, et al. Contamination of natural resources (soil and river water) with *Mycobacterium avium* subspecies *paratuberculosis* in three districts of Uttar Pradesh: A pilot study. *Haryana Vet.* 2012b; 51: 1-5.
138. Singh SV, Vihan VS. Detection of *Mycobacterium avium* subspecies *paratuberculosis* goat milk. *Small Ruminant Research.* 2004; 54: 231-235.

139. Soltani M, Nassiry MR, Shahroudi FE, Bassami MR. Detection of *Mycobacterium paratuberculosis* in Feces and Milk Samples from Holstein Dairy Cows by PCR. *Biotechnology*. 2008; 7; 582-585.
140. Stabel JR, Lambertz A. Efficacy of pasteurization for the inactivation of *Mycobacterium avium* subsp. *Paratuberculosis* in milk. *J food Prot*. 2004; 67 (12):2729-26
141. Stabel JR, Steadham EM, Bolin CA. Heat inactivation of *Mycobacterium paratuberculosis* in raw milk: are current pasteurization conditions effective? *Appl. Environ. Microbiol*. 1997; 63:4975–4977.
142. Stabel JR, Wells SJ, Wagner BA. Relationships between fecal culture, ELISA, and bulk tank milk test results for Johne's disease in US dairy herds. *Journal of Dairy Science*. 2002; 85, 525–531.
143. Steinert M, Birkness K, White E, Fields B, Quinn F. *Mycobacterium avium* bacilli grow saprozoically in coculture with *Acanthamoeba polyphaga* and survive within cyst walls. *Appl Environ Microbiol*. 1998;64(6):2256-61.
144. Stephen R, Schumacher S, Tasara T, Grant IR. Prevalence of *Mycobacterium avium* subspecies *paratuberculosis* in swiss raw milk cheeses collected at the retail level. *J. Dairy Sci*. 2007; 90: 3590-3595.
145. Streeter RN, Hoffsis GF, Bech-Nielsen S, Shulaw WP, Rings DM. Isolation of *Mycobacterium paratuberculosis* from colostrum and milk of subclinically infected cows. *Am. J. Vet. Res*. 1995; 56:1322–1324.
146. Sung N, Collins MT. Thermal tolerance of *Mycobacterium avium* subspecies *paratuberculosis*. *Appl Env Microbiol*. 1998; 64: 999-1005.
147. Sweeney RW, Whitlock RH, Buckley CL, Spencer P, Rosenberger AE, Hutchinson LJ. Diagnosis of paratuberculosis in dairy cattle, using enzyme-linked immunosorbent assay for detection of antibodies against *Mycobacterium paratuberculosis* in milk. *American Journal of Veterinary Research*. 1994; 55: 905–909.
148. Sweeney RW, Whitlock RH, Rosenberger AE. *Mycobacterium paratuberculosis* cultured from milk and supramammary lymph nodes of infected asymptomatic cows. *J. Clin. Microbiol*. 1992a. 30:166–171.
149. Sweeney RW, Whitlock RH, Rosenberger AE. *Mycobacterium paratuberculosis* isolated from fetuses of infected cows not manifesting signs of the disease. *Am. J. Vet. Res*. 1992b; 53:477–480.
150. Szewzik U, Szewzyk R, Manz W, Schleifer KH. Microbiological safety of drinking water. *Annu Rev Microbiol*. 2000; 54; 81-127
151. Taylor TK, Wilks CR, McQueen DS. Isolation of *Mycobacterium paratuberculosis* from the milk of a cow with Johne's disease. *Vet. Rec*. 1981;109:532–533.
152. Thomson R, Tolson C, Carter R, Coulter C, Huygens F, Hargreaves M. Isolation of nontuberculous mycobacteria (ntm) from household water and shower aerosols in patients with pulmonary disease caused by ntm. *Am. J. Clin. Microb*. 2013; 51: 3006–3011.
153. Timms VJ, Daskalopoulos G, Mitchell HM, Neilan BA. The association of *Mycobacterium avium* subsp. *Paratuberculosis* with Inflammatory Bowel Disease. *PLoS ONE*. 2016; 11 (2): e0148731. Doi: 10.1371/journal.pone.0148731
154. Tuci A, Tonon F, Castellani L, Sartini A, Roda G, Marocchi M, et al. Fecal detection of *Mycobacterium avium paratuberculosis* using the IS900 DNA sequence in Crohn's disease and ulcerative colitis patients and healthy subjects. *Dig Dis Sci*. 2011;56:2957–2962.
155. Verma AK, Dhama K, Chakraborty S, Kumar A, Tiwari R, Rahal A, et al. Strategies for combating and eradicating important infectious diseases of animals with particular reference to India: Present and future perspectives. *Asian J. Anim. Vet. Adv*. 2014; 9(2): 77-106. <http://dx.doi.org/10.3923/ajava.2014.77.106>
156. Vilagut L, Pares A, Vinas O, Vila J, Jiménez de Anta MT, Rodés J. Antibodies to mycobacterial 65 kD heat shock protein cross-react with the main mitochondrial antigens in patients with primary biliary cirrhosis. *Eur J Clin Invest*. 1997; 27(8):667–72.10.1046/j.1365-2362.1997.1690724.x
157. Vinodhkumar OR, Gunaseelan L, Ronald BS, Sakhthivalan SM. Slaughterhouse prevalence of

- ovine paratuberculosis in southern India. Trop Anim Health Prod. 2013; 45:1063-1069.
158. Vinodhkumar OR, Gunaseelan L, Sekar M, Ronald BSM, Sakthivelan SM. Association between milk and serum antibodies in naturally infected buffalo (*bubalus bubalis*) paratuberculosis. Buffalo Bulletin. 2014; 33:84-87.
159. Waddell LA, Rajiae A, Sargeant J, Harris J, Amezcua R, Downey L, et al. The zoonotic potential of *Mycobacterium avium* spp. *paratuberculosis*: a systematic review. Can J Public Health. 2008;99:145–55.
160. Wells JE, Bosilevac JM, Kalchayanand N, Arthur TM, Shackelford TL, Wheeler TL, Koohmarie M. Prevalence of *Mycobacterium avium* subsp. *paratuberculosis* in ileocecal Lymph Nodes and on Hides and carcasses from Cull cows and fed Cattle at commercial Beef Processing Plants in the United States. Journal of Food Protection. 2009; 72: 1457-1462.
161. Wendt SL, George KL, Parker BC, Gruft H, Falkinham JO. Epidemiology of infection by nontuberculous mycobacteria. III. Isolation of potentially pathogenic mycobacteria from aerosols. Am. Rev. Respir. Dis. 1980; 122:259-263.
162. Whan L, Ball HJ, Grant IR, Rowe MT. Occurrence of *Mycobacterium avium* subsp. *paratuberculosis* in untreated water in Northern Ireland. Appl Environ Microbiol. 2005;71: 7107–7112.
163. Whiley H, Keegan A, Giglio S, Bentham R. *Mycobacterium avium* complex – the role of potable water in disease transmission. Journal of Applied microbiology. 2012; 113:223-232.
164. Whittington RJ, Marsh IB, Reddacliff LA. Survival of *Mycobacterium avium* subsp. *paratuberculosis* in dam water and sediment. Appl. Environ. Microbiol. 2005; 71:5304–5308.
165. Wilson DJ, Rood K, Biswas P, Byrem TM. Herd-level prevalence of Johne's disease in Utah and adjacent areas of the intermountain west as detected by a bulk-tank milk surveillance project. J Dairy Sci. 2010;93(12):5792-7. doi: 10.3168/jds.2010-3481.
166. Yokota S, Tsubaki S, Kuriyama T, Shimizu H, Ibe M, Mitsuda T, et al. Presence in Kawasaki disease of antibodies to mycobacterial heatshock protein HSP65 and autoantibodies to epitopes of human HSP65 cognate antigen. Clin Immunol Immunopathol. 1993; 67:163–70. 10.1006/clin.1993.1060
167. Yoshimura HH, Graham DY, Estes MK, Merkal RS. Investigation of association of mycobacteria with inflammatory bowel disease by nucleic acid hybridization. J Clin Microbiol. 1987;25:45–51.

#### How to Cite This Article:

Chaubey, K.K., Singh, S.V. (2018). *Mycobacterium avium* subspecies *paratuberculosis* as a Foodborne Pathogen of High Public Health Significance. Indian J. Biotech. Pharm. Res. 6(4):6–21.