

Review Article

Brief review study on citrus fruits: Blessing of nature

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ABSTRACT

The term "citrus" originated from the Latin form of the Greek word 'Kedros', referring to trees such as pine, cedar, and cypress. Citrus is a widely recognized as one of the world's important fruit crops. This genus encompasses flowering trees and shrubs within the Rutaceae family. Citrus fruits are abundant in ascorbic acid and citric acid, contributing to the reinforcement of the immune system by stimulating the production of white blood cells. Additionally, these fruits contain significant levels of macronutrients which include dietary fiber, potassium, folate, thiamine, phosphorus, magnesium, riboflavin, and Vitamin B6. Citrus fruits are also denoted as acid fruits owing to their soluble solids comprised of organic acids and sugar. They are noteworthy for their antioxidants and anti-inflammatory properties. While the seeds of these fruits can generate new trees, they are inedible and considered waste products. The findings indicate that citrus seeds contain various macronutrients, including protein, carbohydrates, vitamins, flavonoids, carotenoids, alkaloids, limonoids, and essential oils. They also demonstrate potential antimicrobial activities, with reports suggesting components such as anticancer (e.g., Taxol), antiviral, chemotherapeutic, and other bioactive constituents, further highlighting citrus fruits' pivotal role as essential components in functional food. This study concludes that the nutraceutical values of citrus fruits also bear implications for agro-industrial waste management, given their distinctive attributes.

KEY WORDS: Valorization, nutraceuticals, antimicrobial, vitamin C, citrus seeds, anticancer effects, antioxidant.

Introduction

The most traded agricultural crop worldwide is citrus, which is the biggest genus in the Rutaceae family. Australia and Southeast Asia are the origins of citrus.(Thakur & Kumari, 2022). With over 160 genera, the Rutaceae family comprises flowering plants. Family members who hold the greatest financial significance include Citrus aurantifolia (lime), Citrus maxima (pomelo), Citrus limeta (sweet lemon), Citrus sinensis (sweet orange), Citrus reticulata (mandarin orange).

(Ben Hsouna et al., 2023) Citrus trees were first cultivated in the tropical and subtropical regions of Asia and the Malay Archipelago; some types are also found in Mediterranean nations. Because of the improved soil and climate here, orange, mandarin, and lemon trees can grow to a higher degree of fruit quality than they could in their original areas..(Ollitrault et al., 2020) The ancestral species of citrus consists of Citrus maxima – Pomelo Citrus 34eticulate – Mandarin orange amongwhichthe important hybrids are Citrus × aurantiifolia – Key lime Citrus × sinensis – Sweet orange.

The main benefit of citrus species is their abundance in vitamin C. Vitamin C has antioxidant qualities and is an antiscorbutic agent. (Yu et al., 2005) Vitamin C and folate, which are essential for maintaining the integrity of immune barriers and supporting the activity of various immune cell types such as phagocytes, natural killer cells, T-cells, and B-

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cells, may be found in citrus fruit juices. As an antioxidant, vitamin C lowers several components of the inflammatory response (Miles & Calder, 2021). For optimal health throughout the life cycle, vitamin C, one of the main antioxidants in circulation, has anti-inflammatory and immune-supporting properties. (Sengupta et al., 2023). During the COVID-19 pandemic, there has been an increase in the proportion of individuals consuming recommended amounts of fruits, including citrus fruits. (Kołota & Głabska, 2021).

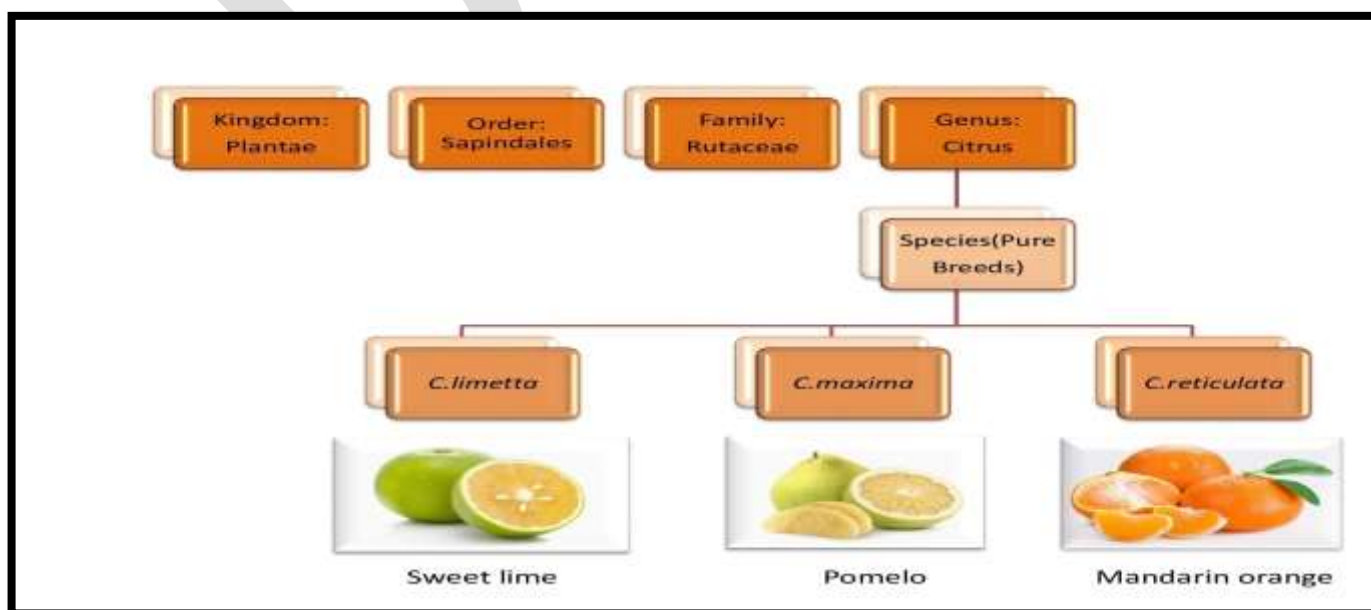
Citrus fruits have become more significant since the COVID-19 pandemic because of their possible antiviral and anti-inflammatory properties, link to a lower chance of developing some malignancies, and positive benefits on cardiovascular and mental health. Citrus fruit juices are high in vitamin C and folate, which help to maintain immunological barriers and promote the activity of numerous immune cells. Vitamin C serves as an antioxidant, reducing certain components of the inflammatory response. Citrus fruit juice polyphenols, such as hesperidin, narirutin, and naringin, exhibit anti-inflammatory properties in model systems and have been demonstrated to lower inflammatory markers in human studies. (Miles & Calder, 2021). The contribution of citrus species in deterrence of life threatening diseases have been assessed (Rafiq et al., 2018) and it has been reported that citrus fruits, citrus fruit extracts and citrus flavonoids exhibit a wide range of promising biological properties due to their phenolic profile and antioxidant properties ((Ramful et al., 2010) Citrus (*Citrus L.* From Rutaceae), contains active phytochemicals

that can protect health. In addition to this, it provides an ample supply of vitamin C, folic acid, potassium and pectin). Studies conducted in vivo and in vitro have demonstrated the potential antidiabetic benefits of citrus flavonoids. It has been discovered that they influence lipid profiles, renal function, and indicators of glycemic management. They also alter 35 metabolic pathways linked to insulin sensitivity and glucose absorption. (Gandhi et al., 2020)

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Over 50 nations in tropical and subtropical climates cultivate citrus for commercial purposes; the total production is 115.52 million tons. With an average productivity of 9.69 tonnes/ha, India ranks as the fourth-largest citrus producer worldwide. (Kumar & Chahal, 2018) After processing, approximately 50% of the fruit is wasted since it cannot be consumed. Fruit pith remains, peels, and seeds fall under the latter category. These wastes include bioactive compounds, thus dumping them directly into the environment can cause serious problems. Disposal in water bodies or seepage into the subsurface water table degrades water quality and endangers aquatic life. The release of these bioactive compounds into open landfills spreads illnesses and emits disagreeable odors (Sharma et al., 2019)

Taxonomical position of the citrus species



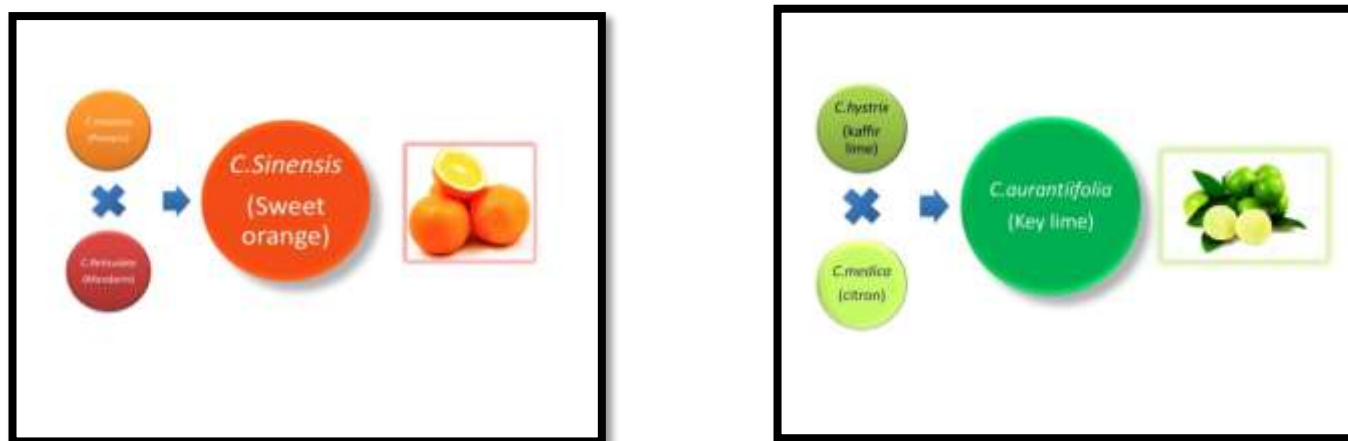


Figure 2: *C. sinensis* and *C. aurantifolia*

Review Literature

The literature review is structured on Antimicrobial, anticancer, and antioxidant activities of the citrus species.

C. aurantifolia

Promising findings have been obtained from studies on *Citrus aurantifolia* seeds' antibacterial properties. Mohammed (2016), (Mohammed & Ayoub, 2016) discovered that the seed extracts in ethanol, methanol, and chloroform all had strong antibacterial activity, with the methanol extract working especially well.

Salih (2015) provided additional support for these results, demonstrating that extracts from *Citrus aurantifolia* impeded the development of harmful bacteria isolated from individuals suffering from sinusitis and asthma. (Salih, 2015) (Mohammed, A. H. Et al., 2016) A zone of inhibition of 19 mm is produced by the aqueous extract against *S. Paratyphi A*, while a zone of inhibition of 13 mm is produced by the chloroform extract against the same bacteria. *S. Typhi* was sensitive to the ethanolic extract of *C. Aurantifolia*, even at the lowest concentration, but resistant to both the aqueous and chloroform extracts of the plant. *S. Paratyphi A* and *S. Paratyphi B* both shown resistance to *C. Aurantifolia* ethanolic extracts.

(Valarmathy et al., 2010) shows that plant extracts from *Citrus aurantifolia* and *Mangifera indica* significantly limit the growth of *Penicillium oxalicum*, *Aspergillus niger*, *Aspergillus terreus*, and *Staphylococcus albus* (Mohammed & Ayoub, 2016) The minimum inhibitory concentration (MIC) ranges for *C. Aurantifolia* water extracts were 12.5 µg/ml to 25 µg/ml, ethanolic extracts were 12.5 µg/ml to 50 µg/ml, and chloroform extracts were 25 µg/ml to 100 µg/ml. The MBC range for both extracts was 25 µg/ml to

100 µg/ml. On the other hand, the MIC and MBC ranges for amoxicillin, the typical antibiotic, were 6.25 µg/ml-12.5 µg/ml and 12.5 µg/ml-25 µg/ml, respectively.

C. limetta

(Shakya et al., 2019) With a maximal zone of inhibition of 15.33 ± 0.577 against *Bacillus* spp., the peel extract of *Citrus limetta* was shown to be more efficient against Gram-positive bacteria than its juice extract. Except for *Klebsiella* on which the peel extract was more effective with a zone of inhibition (10.33 ± 1.527), 10% DMSO showed no zone of inhibition. Among the Gram-negative bacteria, the juice extract of C2 was found to be more effective than its peel extract against the four other bacteria chosen with a maximum zone of inhibition (8.33 ± 2.081) against *E. Coli* ATCC 25922.

C. maxima

(Br Karo et al., 2020) used the disc diffusion method to assess the antibacterial activity of the ethyl acetate fraction of *C. Maxima* peel extract against various microorganisms. These bacterial strains were gram-positive *S. Aureus* and gram-negative *E. Coli*, two species commonly associated with infectious illnesses. The presence or absence of an inhibition zone was used to determine the antibacterial activity of the *C. Maxima* ethyl acetate fraction. The diameter of the inhibition zones surrounding each disk was then measured in 36 petri dishes. The ethyl acetate portion of *C. Maxima* peel extract exhibited mild antibacterial activity in all concentration treatments (25 ppm, 50 ppm, 75 ppm, and 100 ppm) according to the inhibition zone that was obtained. Despite the fact that *S. Aureus* showed the largest zone of inhibition at a dose of 100 ppm, *E. Coli* was at a concentration of 75 ppm.

(Das et al., n.d.) discovered that *Pseudomonas aeruginosa* and *Escherichia coli* were both responsive to *Citrus maxima* (Burm.) Merr.'s ethanolic extract at both concentrations (10 mg/ml & 5 mg/ml). *Pseudomonas aeruginosa* demonstrated the maximum zone of inhibition. *Pseudomonas aeruginosa* had a lower MIC value (0.312 mg/ml) of the extract than *Escherichia coli* (0.625 mg/ml), although both bacteria had the same MBC value (1.25 mg/ml). *Pseudomonas aeruginosa* was the chosen bacteria that responded more strongly to the plant extract than *Escherichia coli*. Given that *Pseudomonas aeruginosa* is the most prevalent gram-negative bacterium that causes both nosocomial and community-acquired illnesses, this indicates the extract's strong antibacterial action. One of the biggest obstacles to the efficient treatment of infections at the moment is the emergence of multidrug-resistant *Pseudomonas aeruginosa*. (Anjum & Mir, 2010).

C. sinensis

(Egbonu & Osuji, 2016) found when compared to the peels (90.21%), the percentage yield of the *C. Sinensis* seeds sample (54.49%) was lower ($p < 0.05$). The orange peels and seeds water extracts demonstrated activity (measured as inhibition zone diameter, IZD in mm) against *Staphylococcus aureus* (14.00 ± 1.00 , 8.67 ± 1.15) and *Escherichia coli* (11.33 ± 1.52 , 9.67 ± 1.15), respectively, with the peels extract eliciting significantly ($P < 0.05$) higher activity. The ethanol extract of orange peels exhibited greater activity (mm) against both *Escherichia coli* (20.00 ± 1.73) and *Staphylococcus aureus* (19.00 ± 1.73), with p-values less than 0.05, in comparison to the seeds (12.33 ± 0.38 , 13.33 ± 1.16).

C. reticulata

Different fractions from *C. reticulata* peel inhibited the growth of gram-positive bacteria at quantities that were less

than necessary for gram-negative bacteria. Of all the bacterial strains, the EtO H soluble fraction was the most effective. Out of all the fractions examined, the acetone extract was shown to be the least effective. For *Staphylococcus aureus* and *Bacillus cereus*, the minimum inhibitory concentration (MIC) in the EtOH-soluble fraction was found to be 300 µg/ml, whereas *B. Coagulans* and *B. Subtilis* required 500 µg/ml to completely block growth. For gram-negative bacteria, *Pseudomonas aeruginosa* and *Escherichia coli* It was discovered that the MIC of the EtO H soluble fraction was 1 2 0 0 and 600 jig/ml, respectively. EtO H soluble fraction's MIC levels were lower than those of the other fractions, most likely as a result of its increased polymethoxylated 37 eticula concentration. (Jayaprakasha et al., 2000).

Anticancer Effects

C. aurantifolia

PLC/PRF/5 liver cancer cells with a p53 mutation may be susceptible to inhibition of cell division and induction of apoptosis by lime peel extract. Limonin, hesperidin, and lime peel extract can all be used at non-toxic doses to prevent PLC/PRF/5 cells from invading. This study also found that the isolated components of limonin and hesperidin had less of an effect in inducing apoptosis than did the lime peel extract. Furthermore, the extract's greater efficacy can be explained by the synergistic action of limonin and hesperidin on apoptosis induction. The anti-cancer effect of the extract is probably due to the combinatorial effects of multiple other compounds found in the metabolome, rather than just the synergistic effect of hesperidin and limonin, as the extract's effect is still greater than that of the combination of the two. (Phucharoenrak et al., 2023).

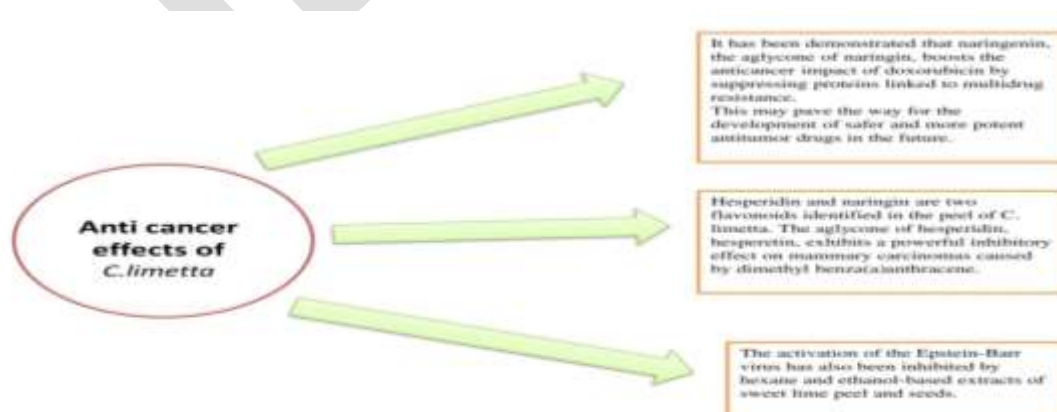


Figure 3: Anticancer effects of *C. limetta*

C.maxima

By raising intracellular ROS levels, CM has demonstrated strong anticancer effects in metastatic breast cancer cells in this study. These results suggested that more research should be done on CM as a potent natural anticancer therapy for breast cancer. Growing research suggests that raising oxidative stress may be a useful tactic for eliminating cancer cells [4]. In breast cancer cells, agents that may raise ROS levels have anticancer properties. (Mursiti et al., 2021).

C.reticulata

This experiment provided a concise summary of peel extract's cytotoxic activity as well as the relationship between total phenolic content. Each cell line's toxicity was

then assessed using its IC50 value. When it comes to colon cancer (HCT) and human breast cancer (MCF7) cell lines, the cytotoxicity of 70% peel extract was typically ineffective since the IC50 was higher than 15. However, the same extract revealed minimal toxicity to normal cells but a significant cytotoxic impact against human hepatocellular liver cancer (HepG2, IC50 9.9 µg/mL). Finally, flavonoids with a high total phenolic content exhibit more potent cytotoxic properties.

Citrus peel extract has been demonstrated to be effective against liver cancer due to its high phenolic content, which includes flavonoids and coumarone compounds.(Ya et al., 2019)

Antioxidant Properties

CITRUS SPECIES	ANTIOXIDANT POTENTIAL
<i>C.maxima</i>	Studies have shown that the fruit juice of <i>Citrus maxima</i> can improve antioxidant status and reduce the risk of oxidative stress (Oyedepo, 2012). This potential is further supported by the presence of phytochemicals, minerals, and vitamins C and E in the fruit juice (Ezeanyikaa, 2022). The leaves of <i>Citrus maxima</i> have also been found to exhibit <i>in vivo</i> antioxidant and hepatoprotective activity (KunduSen, 2011), as well as <i>in vitro</i> antioxidant capacity and <i>in vivo</i> antidiabetic properties (Islam, 2021). These findings collectively suggest that <i>Citrus maxima</i> , particularly its fruit juice and leaves, can be a valuable source of antioxidants.
<i>C.aurantifolia</i>	Research on the antioxidant potential of <i>Citrus aurantifolia</i> , or lime, has consistently shown promising results. Kumari (2013) found that both unripen and ripen lime juices possess strong antioxidant activity, with unripen juices showing higher scavenging activity. This was attributed to the higher levels of ascorbic acid and total phenolic content in unripen juices. Similarly, Namani (2018) reported that lime leaves from Oman contain significant levels of flavonoids and phenolic compounds, which contribute to their moderate antioxidant activity.
	Research has consistently shown the high antioxidant potential of <i>Citrus reticulata</i> . Bocco (1998) and Kamal (2013) both found that the peel and essential oils of this citrus species have significant antioxidant activity. Parhiz (2015) further highlighted the antioxidant and anti-inflammatory properties of the citrus flavonoids hesperidin and hesperetin, which are present in <i>Citrus reticulata</i> . Mishra (2023) specifically identified rutin, a flavonoid found in <i>Citrus</i>

<i>C. reticulata</i>	reticulata, as an effective antioxidant and anti-cancer agent.
<i>C. limetta</i>	Research on the antioxidant potential of Citrus limetta, particularly its juice, leaves, fruits, and peel extracts, has shown promising results. Kumari (2017) found that the juice extract exhibited high antioxidant activity, while PI (2012) and KunduSen (2011) reported effective free radical scavenging and antioxidant activities in the leaves, fruits, and peel extracts. Javed (2013) further supported these findings, demonstrating the antioxidant activity of the essential oil of Citrus limetta var. Mitha. These studies collectively suggest that Citrus limetta possesses significant antioxidant potential.

Conclusion

Citrus species are an easily available treasure trove of mother nature, they contain vitamin C, flavonols, flavonoids, bioactive compounds, limonoids, folic acids, and dietary fibers; thus they can be vividly used ethnopharmacologically. They are used for different types of treatments not only by the medical practitioners also used as home remedies. So, it is highly essential to conduct research study on the different parts of citrus fruits. Valorization of the waste parts of the samples also needs to be enlightened. Overall, this review study sheds light on the protective effects of citrus species. Thinking about the efficacy of the genus, this review paper will draw research attention towards it. It can be categorized as a functional foods because of its high antioxidant activities, also from the different parts of the fruits nutraceuticals can be produced.

References

- Al-Aamri, M. S., Al-Abousi, N. M., Al-Jabri, S. S., Alam, T., & Khan, S. A. (2018). Chemical composition and in-vitro antioxidant and antimicrobial activity of the essential oil of Citrus aurantifolia L. leaves grown in Eastern Oman. Journal of Taibah University Medical Sciences, 13(2), 108–112. <https://doi.org/10.1016/J.JTUMED.2017.12.002>
- Álvarez, R., Carvalho, C. P., Sierra, J., Lara, O., Cardona, D., & Londoño-Londoño, J. (2012). Citrus juice extraction systems: Effect on chemical composition and antioxidant activity of clementine juice. Journal of Agricultural and Food Chemistry, 60(3), 774–781. <https://doi.org/10.1021/jf203353h>
- Amenu, J. D., Neglo, D., & Abaye, D. A. (2019). Comparative Study of the Antioxidant and Antimicrobial Activities of Compounds Isolated from Solvent Extracts of the Roots of <i>Securinegavirosa</i>. Journal of Biosciences and Medicines, 07(08), 27–41. <https://doi.org/10.4236/jbm.2019.78003>
- Anjum, F., & Mir, A. (2010). Susceptibility pattern of pseudomonas aeruginosa against various antibiotics. African Journal of Microbiology Research, 4(10), 1005–1012. <http://www.academicjournals.org/ajmr>
- Aruoma, O. I., Landes, B., Ramful-Baboolall, D., Bourdon, E., Neergheen-Bhujun, V., Wagner, K. H., & Bahorun, T. (2012). Functional benefits of citrus fruits in the management of diabetes. Preventive Medicine, 54(SUPPL.), S12–S16. <https://doi.org/10.1016/J.YPMED.2012.02.012>
- Baker, D. H. A., Ibrahim, E. A., & Salama, Z. A. E.-R. (2021). Citrus Peels as a Source of Bioactive Compounds with Industrial and Therapeutic Applications. Biochemistry. <https://api.semanticscholar.org/CorpusID:245169330>
- ben Hsouna, A., Sadaka, C., Generalić Mekinić, I., Garzoli, S., Švarc-Gajić, J., Rodrigues, F., Morais, S., Moreira, M. M., Ferreira, E., Spigno, G., Brezo-Borjan, T., Akacha, B. ben, Saad, R. ben, Delerue-Matos, C., & Mnif, W. (2023a). The Chemical

- Variability, Nutraceutical Value, and Food-Industry and Cosmetic Applications of Citrus Plants: A Critical Review. In *Antioxidants* (Vol. 12, Issue 2). MDPI. <https://doi.org/10.3390/antiox12020481>
8. ben Hsouna, A., Sadaka, C., Generalić Mekinić, I., Garzoli, S., Švarc-Gajić, J., Rodrigues, F., Morais, S., Moreira, M. M., Ferreira, E., Spigno, G., Brezoborjan, T., Akacha, B. ben, Saad, R. ben, Delermatos, C., & Mnif, W. (2023b). The Chemical Variability, Nutraceutical Value, and Food-Industry and Cosmetic Applications of Citrus Plants: A Critical Review. In *Antioxidants* (Vol. 12, Issue 2). MDPI. <https://doi.org/10.3390/antiox12020481>
 9. Bhatti, S. A., Hussain, M. H., Mohsin, M. Z., Mohsin, A., Zaman, W. Q., Guo, M., Iqbal, M. W., Siddiqui, S. A., Ibrahim, S. A., Ur-Rehman, S., & Korma, S. A. (2022). Evaluation of the antimicrobial effects of Capsicum, Nigella sativa, Musa paradisiaca L., and Citrus limetta: A review. In *Frontiers in Sustainable Food Systems* (Vol. 6). Frontiers Media S.A. <https://doi.org/10.3389/fsufs.2022.1043823>
 10. Br Karo, R. M., Manalu, P., & Simurat, J. P. (2020). Antibacterial Activity of Flavonoid-Rich Fractions Of Citrus Maxima Peel Extract. *Stannum : Jurnal Sains Dan Terapan Kimia*, 2(2), 16–21. <https://doi.org/10.33019/jstk.v2i2.1977>
 11. Chahardehi, A. M., Arsad, H., & Lim, V. (2020). Zebrafish as a successful animal model for screening toxicity of medicinal plants. In *Plants* (Vol. 9, Issue 10, pp. 1–35). MDPI AG. <https://doi.org/10.3390/plants9101345>
 12. Das, S., Borah, M., & Ahmed, S. (n.d.). ANTIBACTERIAL ACTIVITY OF THE ETHANOLIC EXTRACT OF LEAVES OF Citrus maxima (Burm.) Merr. ON ESCHERICHIA COLI AND PSEUDOMONAS AERUGINOSA.
 13. Egbuonu, A., & Osuji, C. (2016). Proximate Compositions and Antibacterial Activity of Citrus sinensis (Sweet Orange) Peel and Seed Extracts. *European Journal of Medicinal Plants*, 12(3), 1–7. <https://doi.org/10.9734/ejmp/2016/24122>
 14. Favela-Hernández, J. M. J., González-Santiago, O., Ramírez-Cabrera, M. A., Esquivel-Ferriño, P. C., & Camacho-Corona, M. D. R. (2016). Chemistry and pharmacology of Citrus sinensis. *Molecules*, 21(2). <https://doi.org/10.3390/molecules21020247>
 15. Freedland, K. E., SKALA, J. A., STEINMEYER, B. C., CARNEY, R. M., & RICH, M. W. (2021). Effects of Depression on Heart Failure Self-Care. *Journal of Cardiac Failure*, 27(5), 522–532. <https://doi.org/10.1016/j.cardfail.2020.12.015>
 16. Garba, S. A., Salisu, B., Yusha'u, M., & Hussain, A. (2016). In vitro Assessment of Antibacterial Activity of Citrus aurantifolia Extracts. *UMYU Journal of Microbiology Research (UJMR)*, 1. <https://doi.org/10.47430/ujmr.1611.001>
 17. Girsang, E., Napiah Nasution, A., & Nyoman Ehrich Lister, I. (n.d.). Comparison Activities of Peel and Extract of Lime (Citrus amblycarpa) as Antioxidant and Antielastase. *American Scientific Research Journal for Engineering*. <http://asrjetsjournal.org/>
 18. Gossiau, A., Li, S., Ho, C. T., Chen, K. Y., & Rawson, N. E. (2011). The importance of natural product characterization in studies of their anti-inflammatory activity. *Molecular Nutrition and Food Research*, 55(1), 74–82. <https://doi.org/10.1002/MNFR.201000455>
 19. Gupta, R., & Sharma, S. (2022). Role of alternatives to antibiotics in mitigating the antimicrobial resistance crisis. *The Indian Journal of Medical Research*, 156, 464–477. https://doi.org/10.4103/ijmr.IJMR_3514_20
 20. Harborne, J. B., & Williams, C. A. (2000). Advances in flavonoid research since 1992. *Phytochemistry*, 55(6), 481–504. [https://doi.org/10.1016/S0031-9422\(00\)00235-1](https://doi.org/10.1016/S0031-9422(00)00235-1)
 21. Jayaprakasha, G. K., Negi, P. S., Sikder, S., Jagan ohanrao, L. M., & Sakariah, K. K. (2000). Antibacterial Activity of Citrus reticulata Peel Extracts (Vol. 55). www.znaturforsch.com
 22. Kołota, A., & Głogowska, D. (2021). Covid-19 pandemic and remote education contributes to improved nutritional behaviors and increased screen time in a polish population-based sample of primary school adolescents: Diet and activity of youth during covid-19 (day-19) study. *Nutrients*, 13(5). <https://doi.org/10.3390/nu13051596>
 23. Kumar, A., & Chahal, T. S. (2018). Studies on diseases and disorders of different citrus germplasm under natural condition. *Journal of Entomology and Zoology Studies*, 6, 406–409. <https://api.semanticscholar.org/CorpusID:89884019>
 24. Kumar, A., Singh Chahal, T., & Adesh Kumar, C. (2018). Studies on diseases and disorders of different citrus germplasm under natural condition. ~ 406 ~

- Journal of Entomology and Zoology Studies, 6(1), 406–409.
25. Kundusen, S., Bala, A., Kar, B., Bhattacharya, S., Mazumder, U. K., Gupta, M., & Haldar, P. K. (2012). Antitumor potential of Citrus limetta fruit peel in Ehrlich ascites carcinoma bearing Swiss albino mice. *Alternative Medicine Studies*, 2(1), 10. <https://doi.org/10.4081/ams.2012.e10>
 26. Lawrence, D. W., Barry, A. L., Richard, O., & C, S. J. (1972). Reliability of the Kirby-Bauer Disc Diffusion Method for Detecting Methicillin-Resistant Strains of Staphylococcus aureus. *Applied Microbiology*, 24(2), 240–247. <https://doi.org/10.1128/am.24.2.240-247.1972>
 27. Mahato, N., Sharma, K., Sinha, M., & Cho, M. H. (2018). Citrus waste derived nutra-/pharmaceuticals for health benefits: Current trends and future perspectives. *Journal of Functional Foods*, 40, 307–316. <https://doi.org/10.1016/J.JFF.2017.11.015>
 28. Mahmoud, A. M., Hernández Bautista, R. J., Sandhu, M. A., & Hussein, O. E. (2019). Beneficial Effects of Citrus Flavonoids on Cardiovascular and Metabolic Health. *Oxidative Medicine and Cellular Longevity*, 2019(1), 5484138. <https://doi.org/https://doi.org/10.1155/2019/5484138>
 29. Mahmoud, M. F., Hamdan, D. I., Wink, M., & El-Shazly, A. M. (2014). Hepatoprotective effect of limonin, a natural limonoid from the seed of Citrus aurantium var. bigaradia, on D-galactosamine-induced liver injury in rats. *Naunyn-Schmiedeberg's Archives of Pharmacology*, 387(3), 251–261. <https://doi.org/10.1007/s00210-013-0937-1>
 30. Manach, C., Regeat, F., Texier, O., Agullo, G., Demigne, C., & Remesy, C. (1996). Bioavailability, metabolism and physiological impact of 4-oxo-flavonoids. *Nutrition Research*, 16(3), 517–544. [https://doi.org/10.1016/0271-5317\(96\)00032-2](https://doi.org/10.1016/0271-5317(96)00032-2)
 31. Manthey, J., Guthrie, N., & Grohmann, K. (2012). Biological Properties of Citrus Flavonoids Pertaining to Cancer and Inflammation. *Current Medicinal Chemistry*, 8(2), 135–153. <https://doi.org/10.2174/0929867013373723>
 32. Miles, E. A., & Calder, P. C. (2021). Effects of Citrus Fruit Juices and Their Bioactive Components on Inflammation and Immunity: A Narrative Review. In *Frontiers in Immunology* (Vol. 12). Frontiers Media S.A. <https://doi.org/10.3389/fimmu.2021.712608>
 33. Mohammed, A. H., Na'inna, S. Z., Yusha'u, M., Salisu, B., Adamu, U., & Garba, S. A. (2016). In vitro Assessment of Antibacterial Activity of Citrus aurantifolia Extracts. *UMYU Journal of Microbiology Research (UJMR)*, 1(1), 1–6. <https://doi.org/10.47430/ujmr.1611.001>
 34. Mohammed, R. M. O., & Ayoub, S. M. H. (2016). Study of Phytochemical Screening and Antimicrobial Activity of <i>Citrus aurantifolia</i> Seed Extracts. *American Journal of Analytical Chemistry*, 07(03), 254–259. <https://doi.org/10.4236/ajac.2016.73022>
 35. Mursiti, S., Amalina, N. D., & Marianti, A. (2021). Inhibition of breast cancer cell development using Citrus maxima extract through increasing levels of Reactive Oxygen Species (ROS). *Journal of Physics: Conference Series*, 1918(5). <https://doi.org/10.1088/1742-6596/1918/5/052005>
 36. Ni, H., Peng, L., Gao, X., Ji, H., Ma, J., Li, Y., & Jiang, S. (2019). Effects of maduramicin on adult zebrafish (Danio rerio): Acute toxicity, tissue damage and oxidative stress. *Ecotoxicology and Environmental Safety*, 168, 249–259. <https://doi.org/10.1016/J.ECOENV.2018.10.040>
 37. Ollitrault, P., Curk, F., & Krueger, R. (2020). Chapter 4 - Citrus taxonomy. In M. Talon, M. Caruso, & F. G. Gmitter (Eds.), *The Genus Citrus* (pp. 57–81). Woodhead Publishing. <https://doi.org/https://doi.org/10.1016/B978-0-12-812163-4.00004-8>
 38. Phucharoenrak, P., Muangnoi, C., & Trachootham, D. (2023). Metabolomic Analysis of Phytochemical Compounds from Ethanolic Extract of Lime (Citrus aurantifolia) Peel and Its Anti-Cancer Effects against Human Hepatocellular Carcinoma Cells. *Molecules*, 28(7). <https://doi.org/10.3390/molecules28072965>
 39. Rafiq, S., Kaul, R., Sofi, S. A., Bashir, N., Nazir, F., & Ahmad Nayik, G. (2018). Citrus peel as a source of functional ingredient: A review. In *Journal of the Saudi Society of Agricultural Sciences* (Vol. 17, Issue 4, pp. 351–358). King Saud University. <https://doi.org/10.1016/j.jssas.2016.07.006>
 40. Ramful, D., Bahorun, T., Bourdon, E., Tarnus, E., & Aruoma, O. I. (2010). Bioactive phenolics and antioxidant propensity of flavedo extracts of Mauritian citrus fruits: Potential prophylactic ingredients for functional foods application. *Toxicology*, 278(1), 75–

87. <https://doi.org/https://doi.org/10.1016/j.tox.2010.01.012>
41. Rath, D., Kar, B., Pattnaik, G., & Bhukta, P. (2023). Synergistic Effect of Naringin and Glimepiride in Streptozotocin-induced Diabetic Rats. *Current Diabetes Reviews*, 20. <https://doi.org/10.2174/1573399820666230817154835>
 42. Rawson, N. E., Ho, C.-T., & Li, S. (2014). Efficacious anti-cancer property of flavonoids from citrus peels. *Food Science and Human Wellness*, 3(3–4), 104–109. <https://doi.org/10.1016/J.FSHW.2014.11.001>
 43. Roquia, A., Mustafa Al-Lawati, W., Al-Hadrami, I. S., Al-Abdali, S. S., Rahman Al-Rabaani, M. A., & Gopal, G. (2022). Biogenic Synthesis, Characterization and Antimicrobial Activity of Silver Nanoparticles Mediated by Oil Extracted from Waste Seeds of Citrus Sinensis. *Nano LIFE*, 13(01), 2350003. <https://doi.org/10.1142/S1793984423500034>
 44. Salih, N. (2015). Evaluation of the Antimicrobial Effects of Citrus Aurantifolia (Key Lime) Against Different Microbial Species Isolated From Asthma and Sinusitis Patients. *WORLD JOURNAL OF PHARMACY AND PHARMACEUTICAL SCIENCES*, 4, 324–334.
 45. Seid, L., Lakhdari, D., Belgherbi, O., Laourari, I., Golzadeh, N., & Chouder, D. (2023). Valorization of citrus sinensis wastes for the production of orange essential oil: identification of chemical composition and antibacterial activity. *Journal of Fundamental and Applied Sciences*, 14(3), 490–508. <https://doi.org/10.4314/jfas.1252>
 46. Sengupta, S., Banerjee, S., Nayek, S. N., Das, P., Chakraborty, D., Mukherjee, A., Mandal, T., Majumder, A., Saha, M., Chatterjee, S., & Bhattacharya, M. (2023). A brief comparative study of the natural sources (lemons) in the basis of protein, vitamin C, their antibacterial, anthelmintic and cell viability on immune cells. *International Journal of Herbal Medicine*, 11(5), 14–21. <https://doi.org/10.22271/flora.2023.v11.i5a.883>
 47. Sengupta, S., Saha, M., Ghosh, N., Bhattacharya, M., Chatterjee, S., & Ghosh, R. (2023a). Green synthesis of gold nano-conjugates using commonly used citrus species and evaluation of its In- vitro antibacterial efficacy against *Staphylococcus aureus*: A comparative study. *International Journal of Herbal Medicine*, 11, 38–43. <https://doi.org/10.22271/flora.2023.v11.i2a.858>
 48. Sengupta, S., Saha, M., Ghosh, N., Bhattacharya, M., Chatterjee, S., & Ghosh, R. (2023b). Green synthesis of gold nano-conjugates using commonly used citrus species and evaluation of its In- vitro antibacterial efficacy against *Staphylococcus aureus*: A comparative study. *International Journal of Herbal Medicine*, 11, 38–43. <https://doi.org/10.22271/flora.2023.v11.i2a.858>
 49. Shakya, A., Luitel, B., Kumari, P., Devkota, R., Dahal, P. R., & Chaudhary, R. (2019a). Comparative Study of Antibacterial Activity of Juice and Peel Extract of Citrus Fruits. *Tribhuvan University Journal of Microbiology*, 6, 82–88. <https://doi.org/10.3126/tujm.v6i0.26589>
 50. Shakya, A., Luitel, B., Kumari, P., Devkota, R., Dahal, P. R., & Chaudhary, R. (2019b). Comparative Study of Antibacterial Activity of Juice and Peel Extract of Citrus Fruits. *Tribhuvan University Journal of Microbiology*, 6(0), 82–88. <https://doi.org/10.3126/tujm.v6i0.26589>
 51. Shakya, A., Luitel, B., Kumari, P., Devkota, R., Dahal, P. R., & Chaudhary, R. (2019c). Comparative Study of Antibacterial Activity of Juice and Peel Extract of Citrus Fruits. *Tribhuvan University Journal of Microbiology*, 6, 82–88. <https://doi.org/10.3126/tujm.v6i0.26589>
 52. Sharma, K., Mahato, N., & Lee, Y. R. (2019a). Extraction, characterization and biological activity of citrus flavonoids. *Reviews in Chemical Engineering*, 35(2), 265–284. <https://doi.org/10.1515/revce-2017-0027>
 53. Sharma, K., Mahato, N., & Lee, Y. R. (2019b). Extraction, characterization and biological activity of citrus flavonoids. 35(2), 265–284. <https://doi.org/doi:10.1515/revce-2017-0027>
 54. Suri, S., Singh, A., & Nema, P. K. (2022). Current applications of citrus fruit processing waste: A scientific outlook. *Applied Food Research*, 2(1), 100050. <https://doi.org/10.1016/J.AFRES.2022.100050>
 55. Thakur, N., & Kumari, S. (2022a). Preliminary screening of phytochemicals and antimicrobial activity of *Citrus pseudolimon*. *Advances in Traditional Medicine*, 22(2), 425–435. <https://doi.org/10.1007/s13596-021-00561-y>

56. Thakur, N., & Kumari, S. (2022b). Preliminary screening of phytochemicals and antimicrobial activity of Citrus pseudolimon. *Advances in Traditional Medicine*, 22(2), 425–435. <https://doi.org/10.1007/s13596-021-00561-y>
57. Tung, Y.-C., Chang, W.-T., Li, S., Wu, J.-C., Badmeav, V., Ho, C.-T., & Pan, M.-H. (2018). Citrus peel extracts attenuated obesity and modulated gut microbiota in mice with high-fat diet-induced obesity. *Food & Function*, 9(6), 3363–3373. <https://doi.org/10.1039/C7FO02066J>
58. Valarmathy, K., Gokulakrishnan, M., Salma Kausar, M., & Paul, K. (2010). A study of antimicrobial activity of ethanolic extracts of various plant leaves against selected microbial species. In *Article in International Journal of Pharma Sciences and Research* (Vol. 1, Issue 8). <https://www.researchgate.net/publication/50434390>
59. Vasudeva, N., & Sharma, T. (2012). Chemical Composition and Antimicrobial Activity of Essential Oil of Citrus limettioides Tanaka. *Journal of Pharmaceutical Technology and Drug Research*, 1, 2. <https://doi.org/10.7243/2050-120X-1-2>
60. Viswanatha, G. L., Shylaja, H., & Moolemath, Y. (2017). The beneficial role of Naringin- a citrus bioflavonoid, against oxidative stress-induced neurobehavioral disorders and cognitive dysfunction in rodents: A systematic review and meta-analysis. *Biomedicine & Pharmacotherapy*, 94, 909–929. <https://doi.org/https://doi.org/10.1016/j.biopha.2017.07.072>
61. Wang, H., Tan, J. T. T., Emelyanov, A., Korzh, V., & Gong, Z. (2005). Hepatic and extrahepatic expression of vitellogenin genes in the zebrafish, *Danio rerio*. *Gene*, 356(1–2), 91–100. <https://doi.org/10.1016/j.gene.2005.03.041>
62. Xue, J.-C., Yuan, S., Meng, H., Hou, X.-T., Li, J., Zhang, H.-M., Chen, L.-L., Zhang, C.-H., & Zhang, Q.-G. (2023). The role and mechanism of flavonoid herbal natural products in ulcerative colitis. *Biomedicine & Pharmacotherapy*, 158, 114086. <https://doi.org/https://doi.org/10.1016/j.biopha.2022.114086>
63. Ya, S., Mohamed H, & Sy, H. (2019). Citation: Selim YA, Mohamed H and Hussien SY. Efficacious Anti-Cancer Property of Liver from Peels Extract of Egyptian Citrus reticulata Efficacious Anti-cancer Property of Liver from Peels Extract of Egyptian Citrus reticulata. *Austin J Nutri Food Sci*, 7(3). www.austinpublishinggroup.com
64. Yu, J., Wang, L., Walzem, R. L., Miller, E. G., Pike, L. M., & Patil, B. S. (2005). Antioxidant activity of citrus limonoids, flavonoids, and coumarins. *Journal of Agricultural and Food Chemistry*, 53(6), 2009–2014. <https://doi.org/10.1021/jf0484632>
65. Zhang, C., Willett, C., & Fremgen, T. (2003). Zebrafish: An Animal Model for Toxicological Studies. *Current Protocols in Toxicology*, 17(1), 1.7.1-1.7.18. <https://doi.org/https://doi.org/10.1002/0471140856.tx0107s17>
66. Zou, Z., Xi, W., Hu, Y., Nie, C., & Zhou, Z. (2016). Antioxidant activity of Citrus fruits. *Food Chemistry*, 196, 885–896. <https://doi.org/https://doi.org/10.1016/j.foodchem.2015.09.072>

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